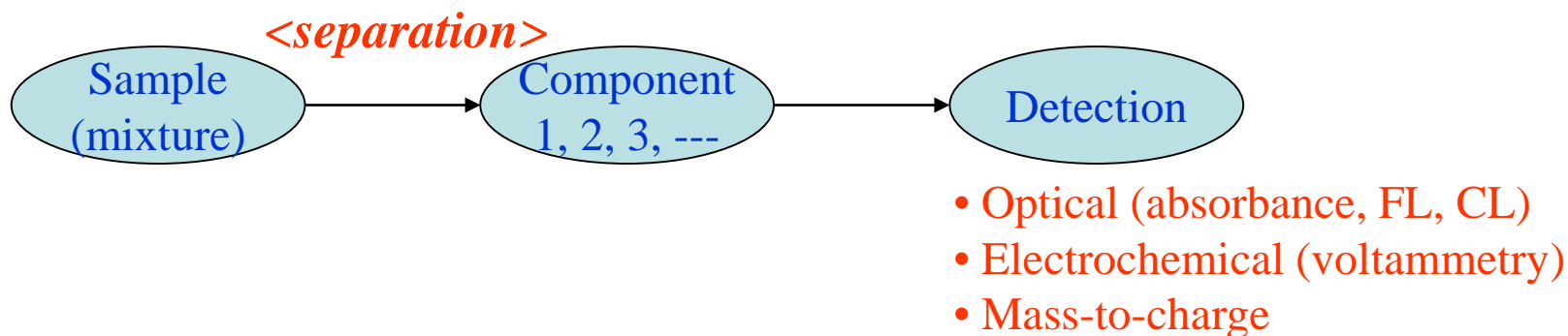


**Chapter 26:**  
**Introduction to Chromatographic Separations**

In real analytical problems,  
we must identify and quantitate one or more components from a complex mixture

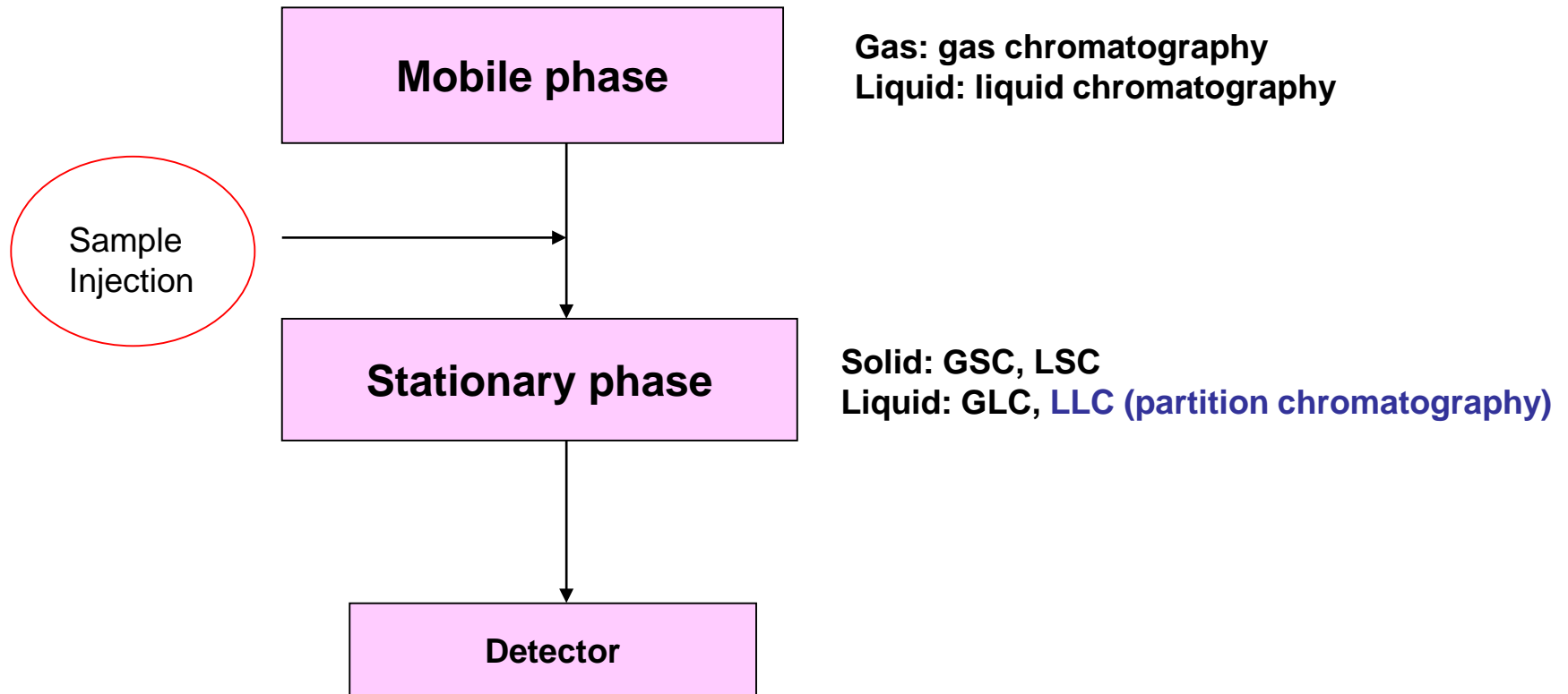
*Separation of mixture into each component is the first step in analysis*



# What is Chromatography?

**Martin and Synges: Nobel Prize in 1952**

-Chromatography operates on the same principle as extraction,  
but **one phase is held in place while the other moves past it.**



**TABLE 26-1** Classification of Column Chromatographic Methods

General Classification	Specific Method	Stationary Phase	Type of Equilibrium
1. Gas chromatography (GC)	a. Gas-liquid chromatography (GLC)	Liquid adsorbed or bonded to a solid surface	Partition between gas and liquid
	b. Gas-solid	Solid	Adsorption
2. Liquid chromatography (LC)	a. Liquid-liquid, or partition	Liquid adsorbed or bonded to a solid surface	Partition between immiscible liquids
	b. Liquid-solid, or adsorption	Solid	Adsorption
	c. Ion exchange	Ion-exchange resin	Ion exchange
	d. Size exclusion	Liquid in interstices of a polymeric solid	Partition/sieving
	e. Affinity	Group specific liquid bonded to a solid surface	Partition between surface liquid and mobile liquid
3. Supercritical fluid chromatography (SFC; mobile phase: supercritical fluid)		Organic species bonded to a solid surface	Partition between supercritical fluid and bonded surface

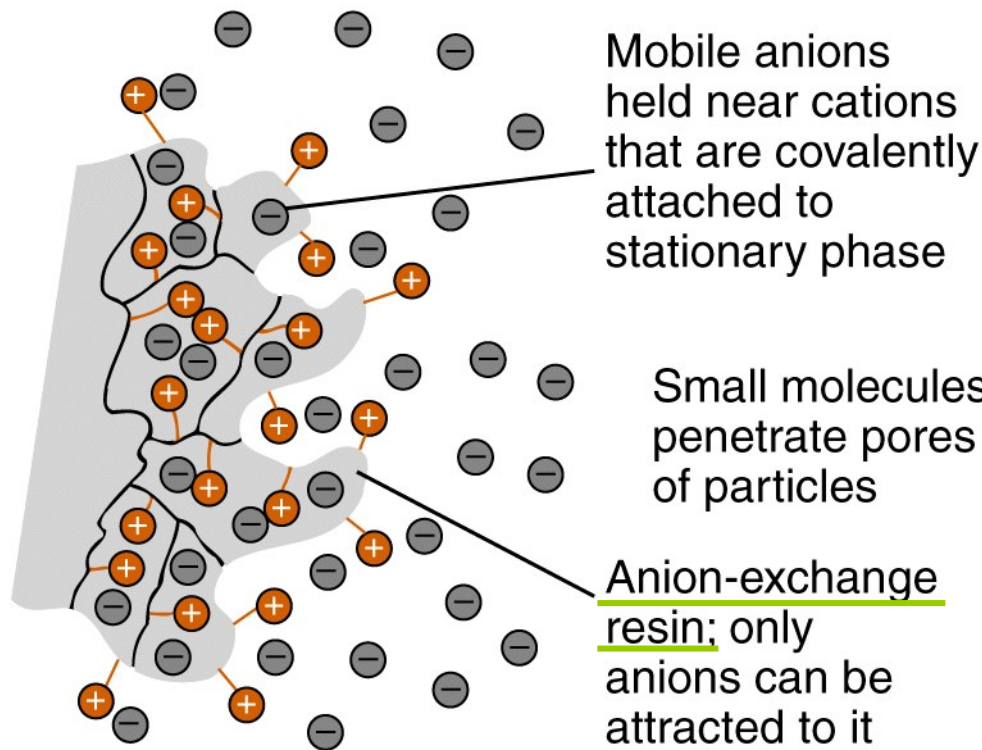
## Ion-exchange chromatography

Stat. phase:

-  $\text{SO}_3^-$

-  $\text{N}(\text{CH}_3)_3^+$

Mobile phase: liquid



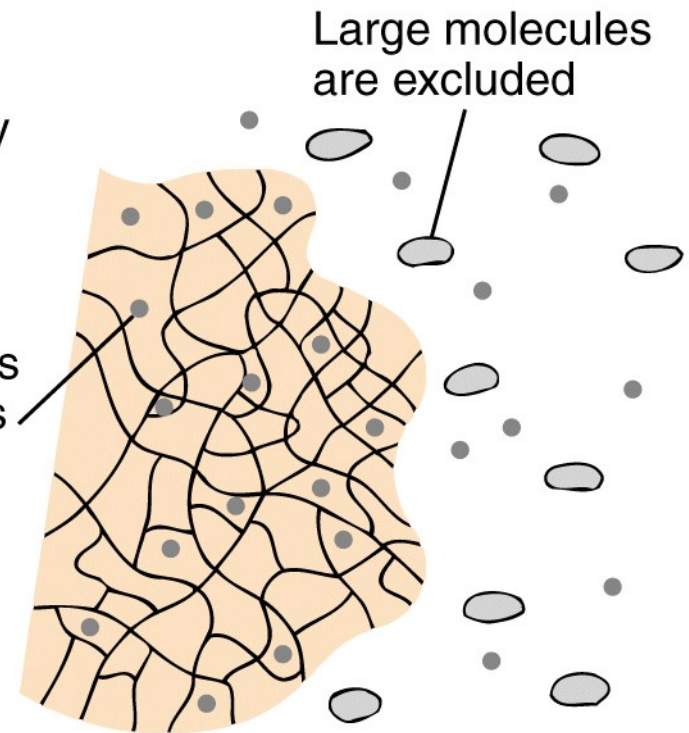
Ion-exchange chromatography

## Molecular exclusion chromatography

: Gel permeation c. (GPC), Gel filtration C

Stat. phase: gel

Mobile phase: liquid



Molecular exclusion chromatography

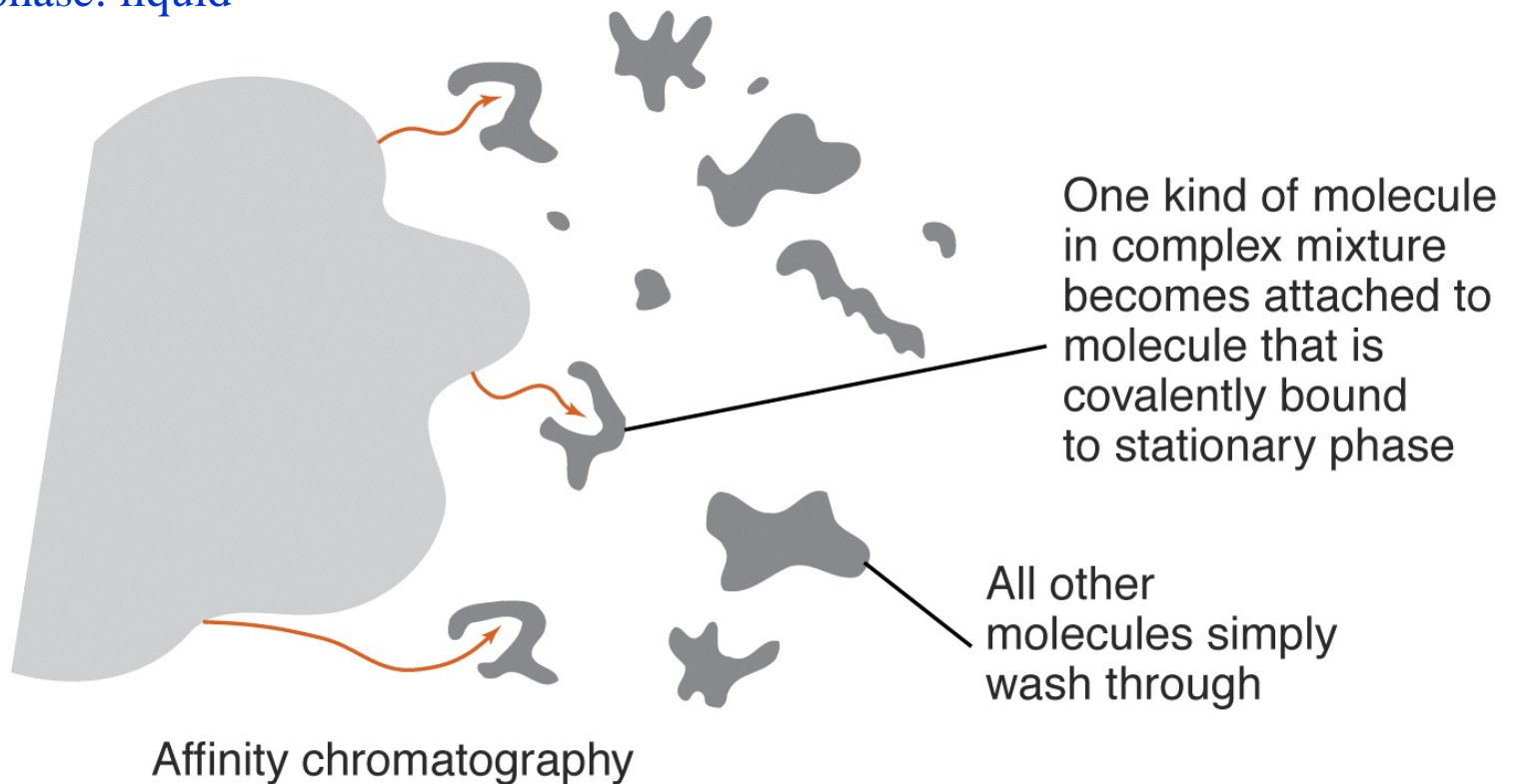
## Affinity chromatography

Stat. phase:

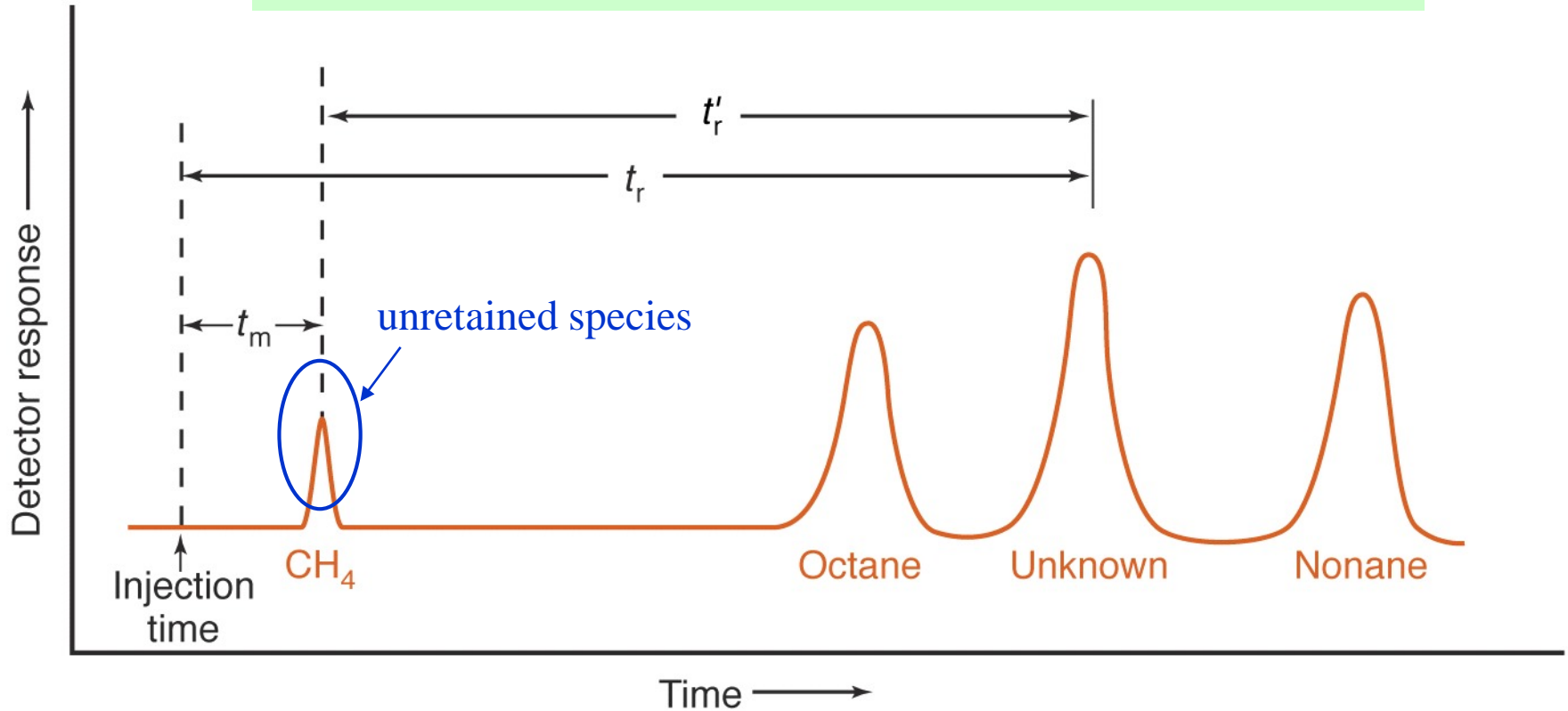
- Immobilized antibody
- Specific protein

Mobile phase: liquid

The most selective kind of chromatography



# Chromatogram



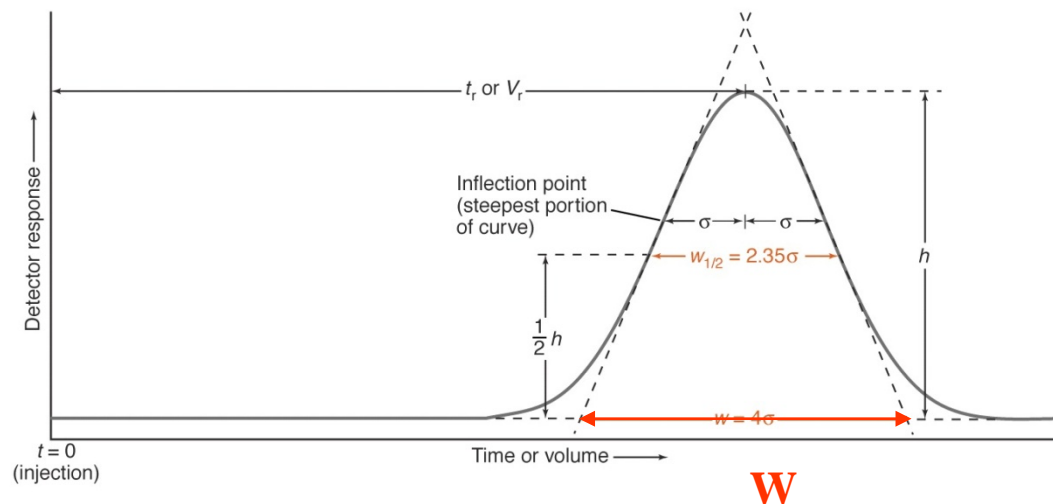
- **Retention time** for each component:  $t_r$
- **Dead time** for unretained species:  $t_m$
- **Adjusted retention time** ( $t_r'$ ) =  $t_r - t_m$
- **Capacity factor** ( $k'$ ) =  $(t_r - t_m)/t_m = t_r'/t_m$
- **Relative retention** ( $\alpha$ ) for any two components (A, B) =  $(t_r')_B / (t_r')_A = (k')_B / (k')_A$   
Selectivity factor  $= K_B / K_A$  (partition coefficient)

# Efficiency of Separation

Two factors to determine separation efficiency:

- **Difference in elution time between peaks** (larger time difference → better separation)
- **Broadness of the separated peaks** (the wider the peaks → the poorer their separation)
- **Resolution** of two peaks from each other is defined as ( $R$ ,  $R_s$ )

$$R_s = (\text{distance between peaks})/(\text{average base width})$$
$$= (\Delta t_r)/[(W_1 + W_2)/2] \approx (\Delta t_r)/W \quad (\text{in most case, } W_1 \approx W_2)$$



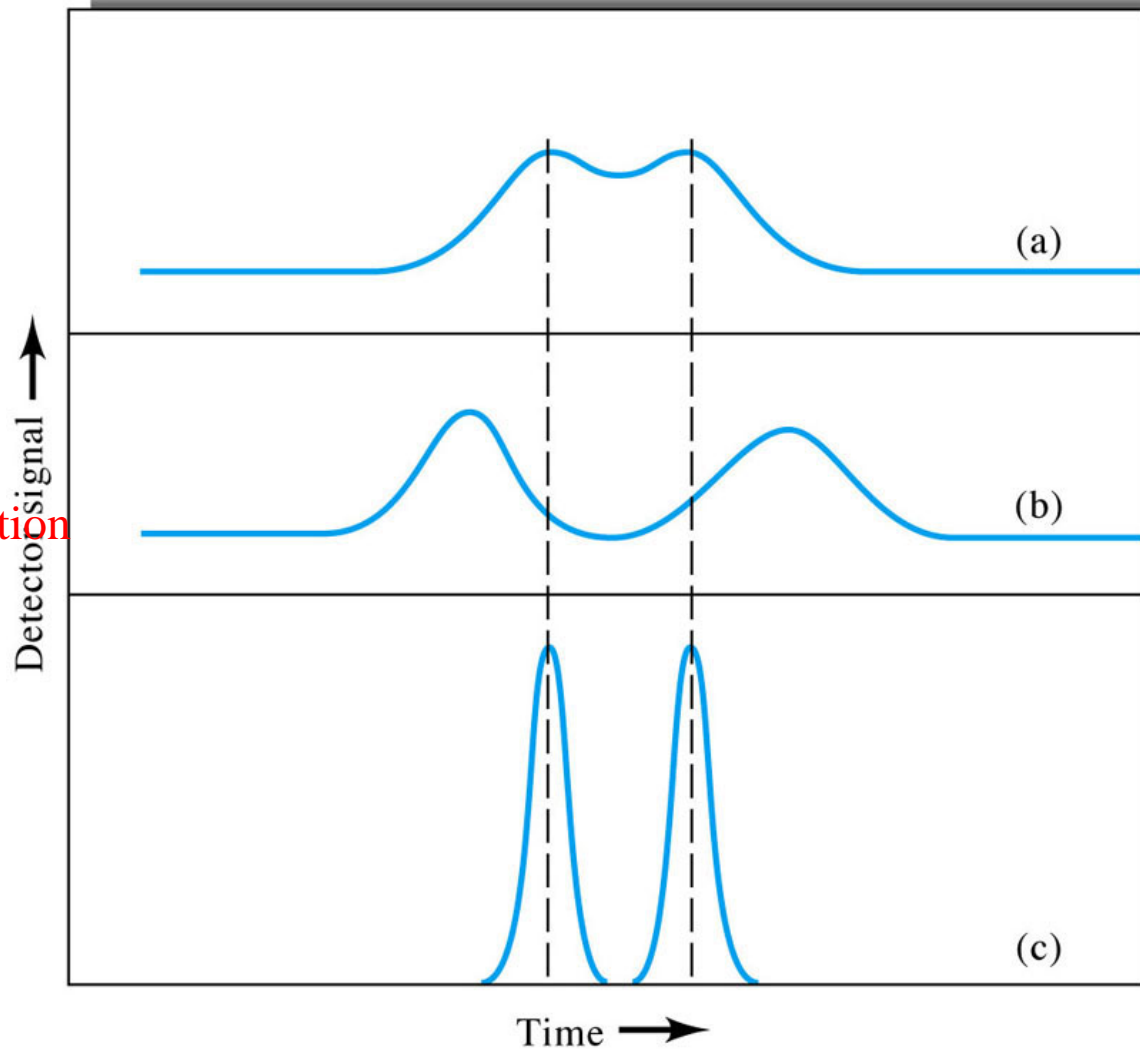


# Efficiency of Separation

original

Increase in  
band separation

Decrease in  
band width



# Gas Chromatography

**Mobile phase (carrier gas): gas (He, N<sub>2</sub>, H<sub>2</sub>)**

- do not interact with analytes
- only transport the analyte through the column

**Analyte: volatile liquid or gas**

**Stationary phase:**

- solid (GSC) or non-volatile liquid (GLC)

**GSC (gas-solid adsorption chromatography)**

- semi-permanent retention of active or polar molecules
- severe tailing of elution peaks

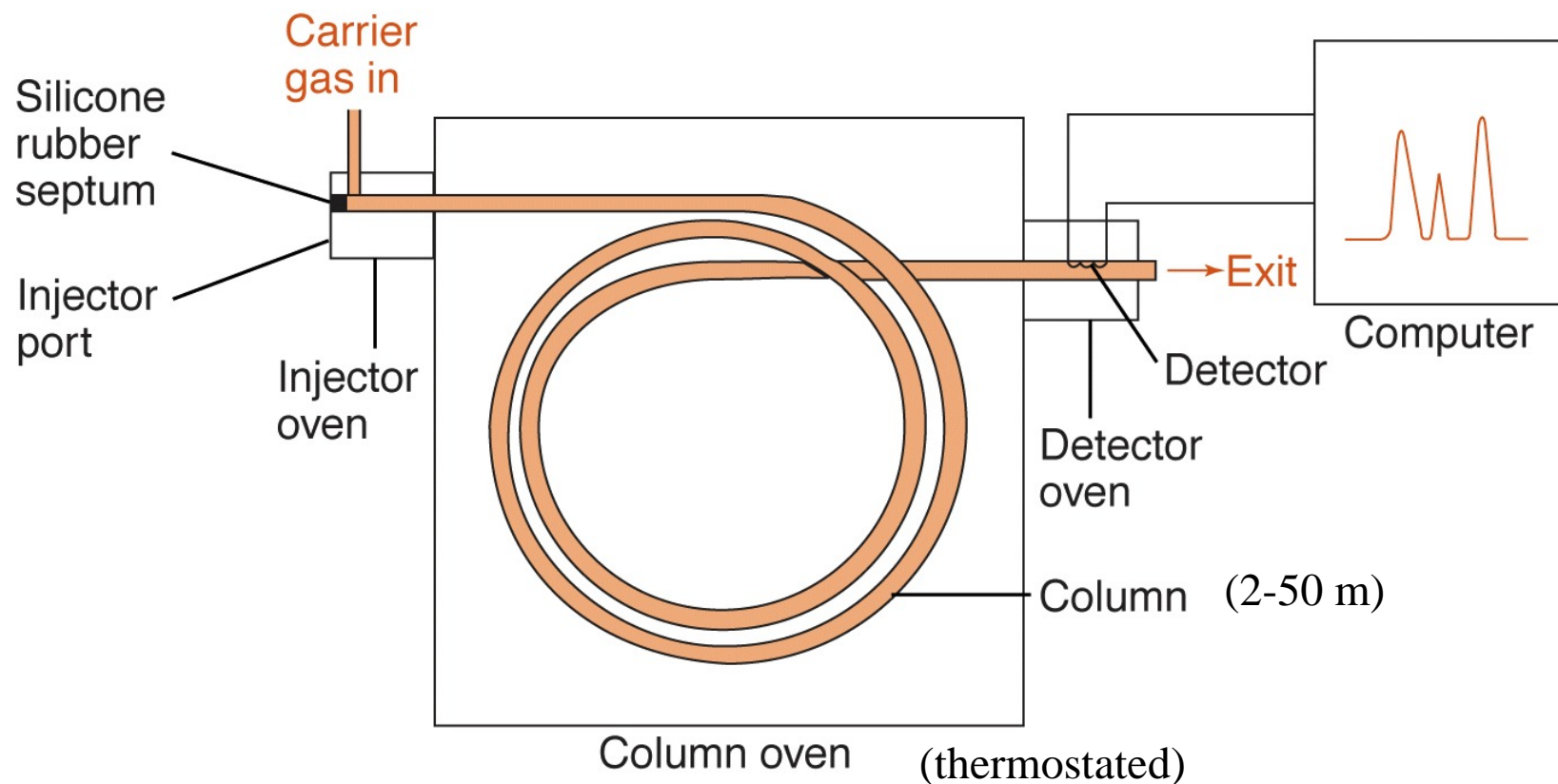
**GLC (gas-liquid partition chromatography)**

- non-volatile liquid is coated on the inside of the column or on a fine solid support
- In 1955, the first commercial apparatus for GLC appeared on the market

# Instrument for gas chromatography

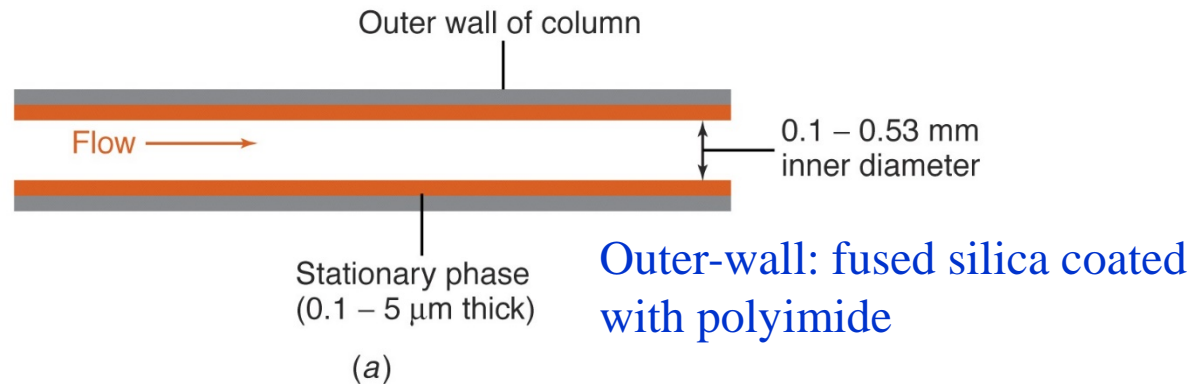
Temp of a sample injector port:

50 °C above the b.p. of least volatile component of the sample → rapidly evaporates



The column should be hot enough to provide sufficient vapor pressure for analyte to be eluted in a reasonable time.

# Open Tubular Columns



- Higher  $N$ , smaller  $H$  → Higher resolution  
( $H = A + B/U + CU$ , no  $A$  term in OTC)
- Higher flow rate → shorter analysis time

Thin coating: small  $C$ -term (decreased  $H$ ) :

Compared with packed columns,

OTC offers higher resolution, shorter analysis time, greater sensitivity, lower sample capacity

# Liquid Sta. Phase

Choice of liquid phase for a given problem:

“like dissolves like”

- **Nonpolar** columns: best for **nonpolar** solutes

- Polar columns for polar solutes

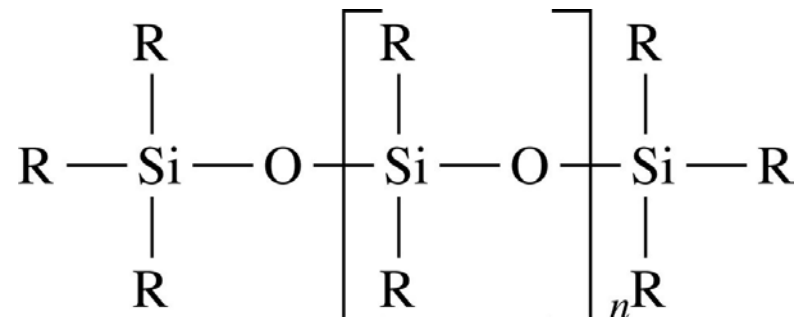
- As a column ages,

→ stationary phases bakes off

→ surface silanol groups (Si-OH) are exposed

→ peak tailing (polar analyte)

Therefore, stationary phase is covalently attached to silica surface

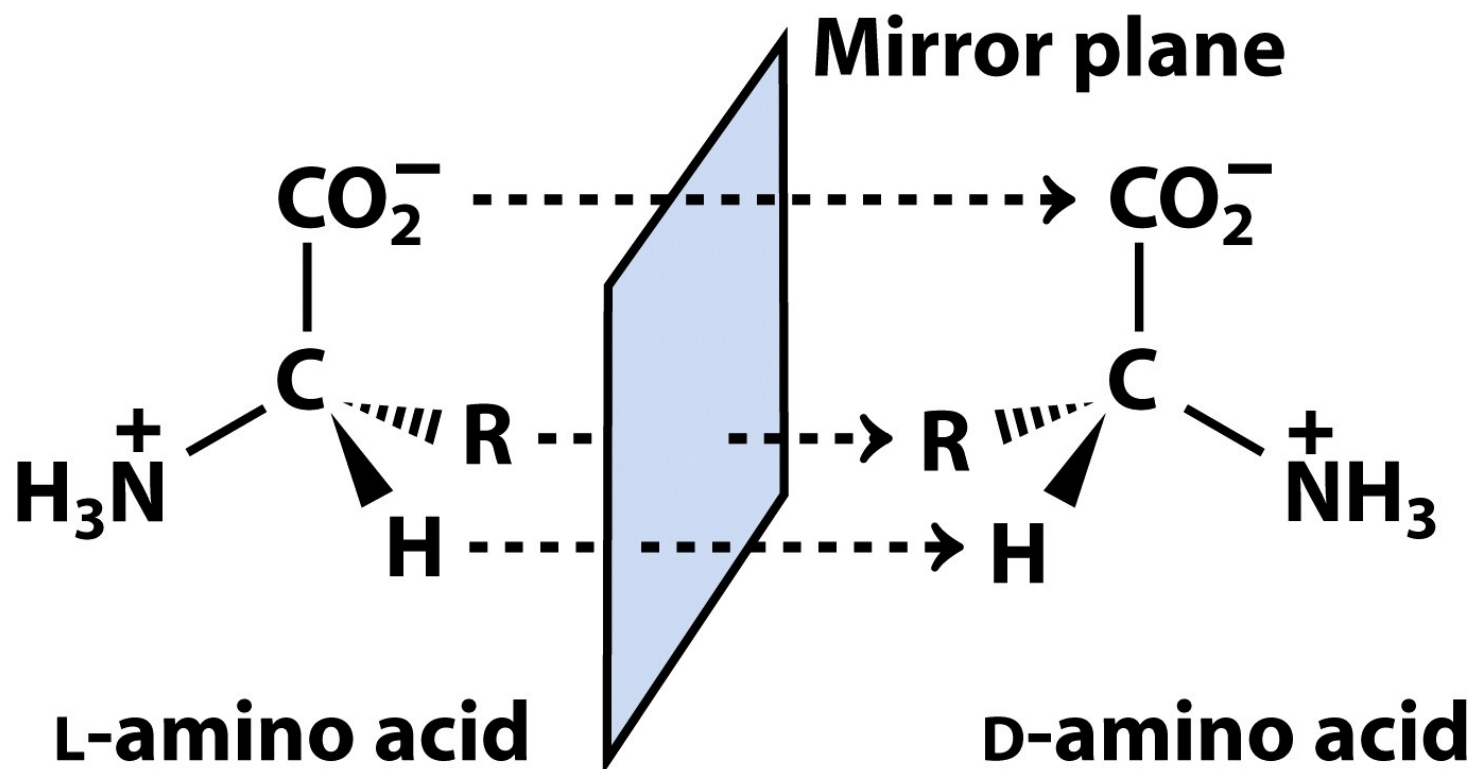


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TABLE 23-1 Common stationary phases in capillary gas chromatography

Structure	Polarity	Temperature range
$\left[ \begin{array}{c} \text{C}_6\text{H}_5 \\   \\ \text{O}-\text{Si} \\   \\ \text{C}_6\text{H}_5 \end{array} \right]_x \left[ \begin{array}{c} \text{CH}_3 \\   \\ \text{O}-\text{Si} \\   \\ \text{CH}_3 \end{array} \right]_{1-x}$ <p>(Diphenyl)<sub>x</sub>(dimethyl)<sub>1-x</sub> polysiloxane</p>	$x = 0$ Nonpolar $x = 0.05$ Nonpolar $x = 0.35$ Intermediate polarity $x = 0.65$ Intermediate polarity	$-60^\circ$ – $320^\circ$ $-60^\circ$ – $320^\circ$ $0^\circ$ – $300^\circ$ $50^\circ$ – $370^\circ$
$\left[ \begin{array}{c} \text{CN} \\   \\ \text{O}-\text{Si} \\   \\ \text{C}_6\text{H}_5 \end{array} \right]_{0.14} \left[ \begin{array}{c} \text{CH}_3 \\   \\ \text{O}-\text{Si} \\   \\ \text{CH}_3 \end{array} \right]_{0.86}$ <p>(Cyanopropylphenyl)<sub>0.14</sub>(dimethyl)<sub>0.86</sub> polysiloxane</p>	Intermediate polarity	$-20^\circ$ – $280^\circ$
$\left[ \text{CH}_2\text{CH}_2\text{O} \right]_n$ <p>Carbowax (poly(ethylene glycol))</p>	Strongly polar	$40^\circ$ – $250^\circ$
$\left[ \begin{array}{c} \text{CN} \\   \\ \text{O}-\text{Si} \\   \\ \text{CN} \end{array} \right]_{0.9} \left[ \begin{array}{c} \text{CN} \\   \\ \text{O}-\text{Si} \\   \\ \text{C}_6\text{H}_5 \end{array} \right]_{0.1}$ <p>(Biscyanopropyl)<sub>0.9</sub>(cyanopropylphenyl)<sub>0.1</sub> polysiloxane</p>	Strongly polar	$0^\circ$ – $275^\circ$

# Chiral Separation



# Quantitative and Qualitative Analysis by GC

## Qualitative analysis:

- retention time (GC-FID, TCD, ECD...): comparison with authentic sample
- mass (GC-MS)

## Quantitative analysis:

- peak area or peak height

**TABLE 27-1** Typical Gas Chromatographic Detectors

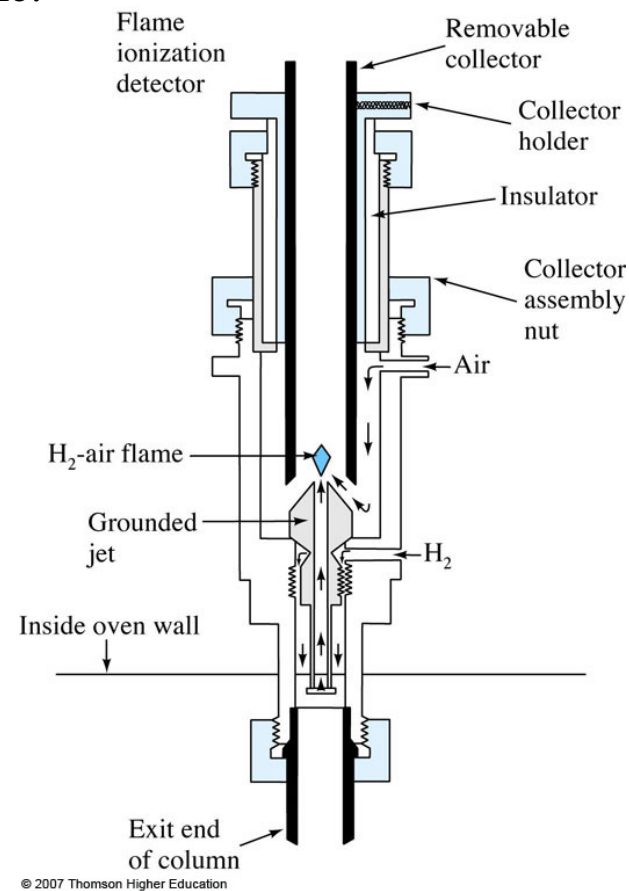
Type	Applicable Samples	Typical Detection Limit
Flame ionization	Hydrocarbons	1 pg/s
Thermal conductivity	Universal detector	500 pg/mL
Electron capture	Halogenated compounds	5 fg/s
Mass spectrometer (MS)	Tunable for any species	0.25 to 100 pg
Thermionic	Nitrogen and phosphorous compounds	0.1 pg/s (P), 1 pg/s (N)
Electrolytic conductivity (Hall)	Compounds containing halogens, sulfur, or nitrogen	0.5 pg Cl/s, 2 pg S/s, 4 pg N/s
Photoionization	Compounds ionized by UV radiation	2 pg C/s
Fourier transform IR (FTIR)	Organic compounds	0.2 to 40 ng

# Flame Ionization Detector (FID)

- Most widely used and generally applicable detector
- Column eluate is mixed with  $\text{H}_2/\text{air}$  and then burned in flame.



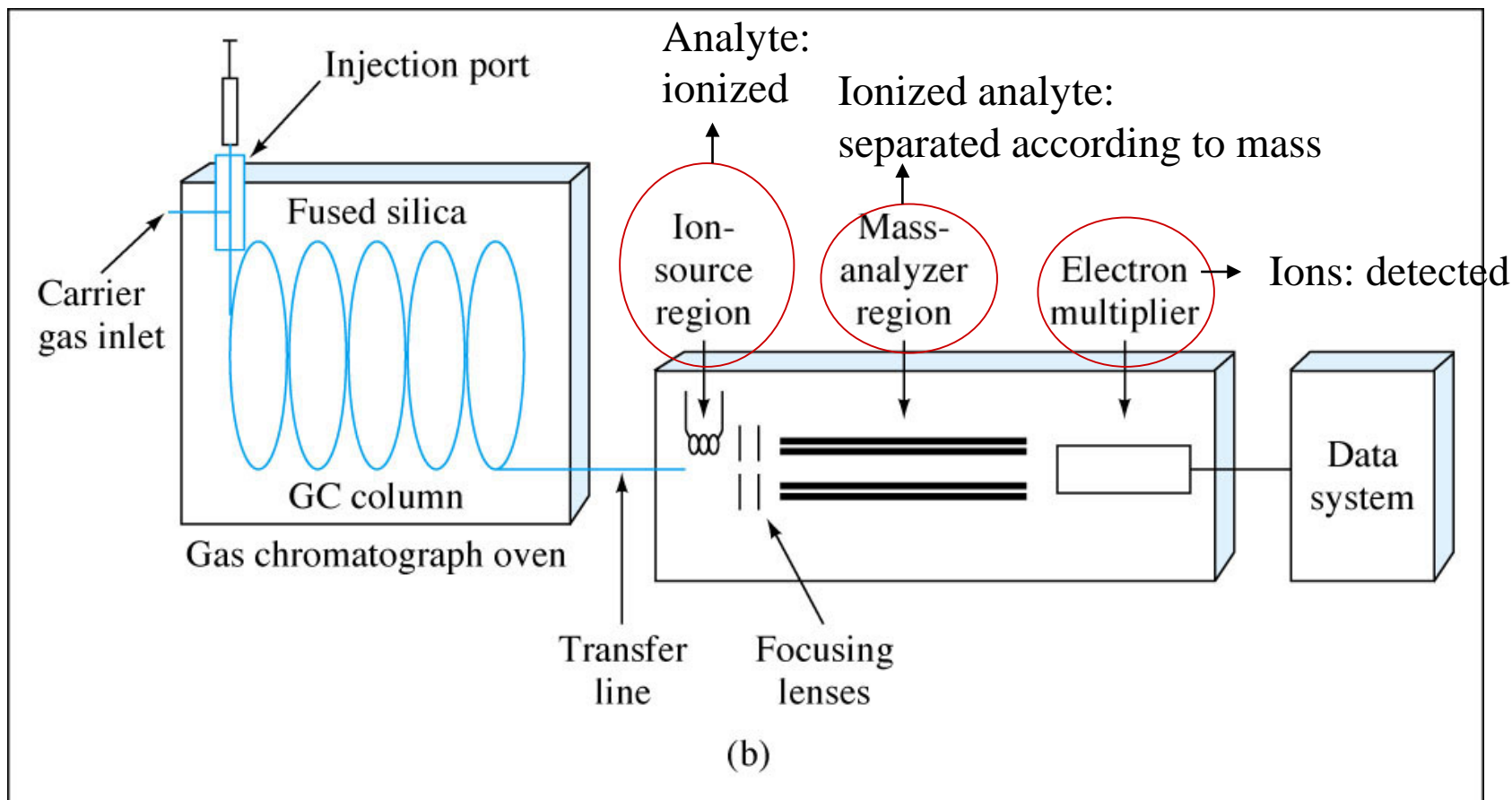
- Most organic compounds, when pyrolyzed at the temp of  $\text{H}_2/\text{air}$  flame, produce ions ( $\sim 1/10^5$ ) and electrons that can conduct electricity through the flame
- FID responds to the # carbons entering the detector per unit time  
: **mass sensitive rather than concentration-sensitive**
- Functional groups (carbonyl, carboxyl, halogen) yield fewer ions
- FID is insensitive to noncombustible gases ( $\text{H}_2\text{O}$ ,  $\text{CO}_2$ ,  $\text{O}_2$ ,  $\text{N}_2$ ,  $\text{SO}_2$ , and  $\text{NO}_x$ )



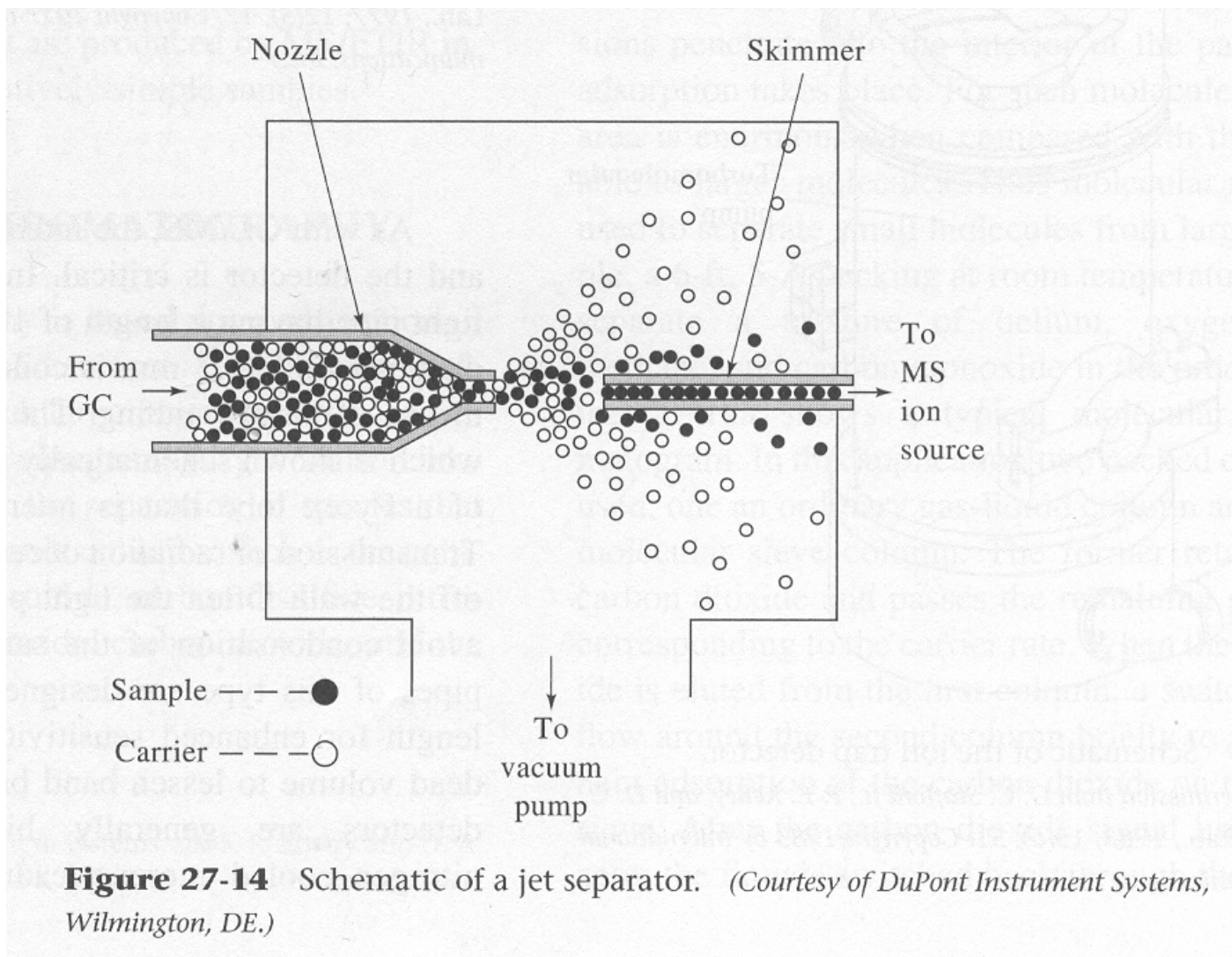


# GC-Mass Spectrometer

Mass spectrometer is a powerful detector for both **qualitative and quantitative analysis** of analyte in gas or liquid chromatography



# GC-Mass Spectrometer



# HPLC

**Mobile phase: liquid**

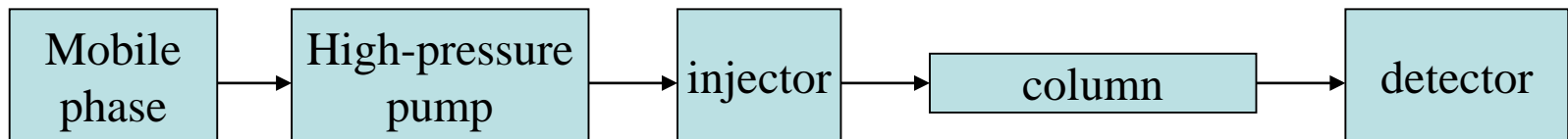
**Analyte: non-volatile liquid**

**Stationary phase:**

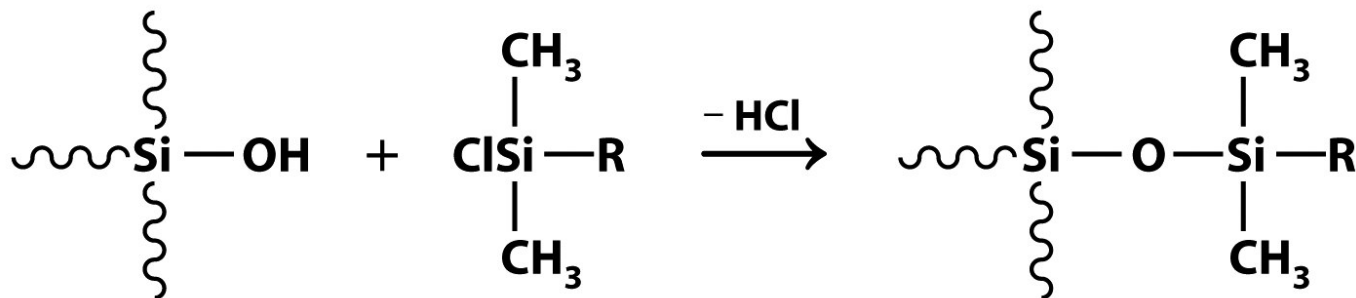
- solid (GSC) or non-volatile liquid (GLC)

HPLC; uses high-pressure pump to deliver liquid mobile phase

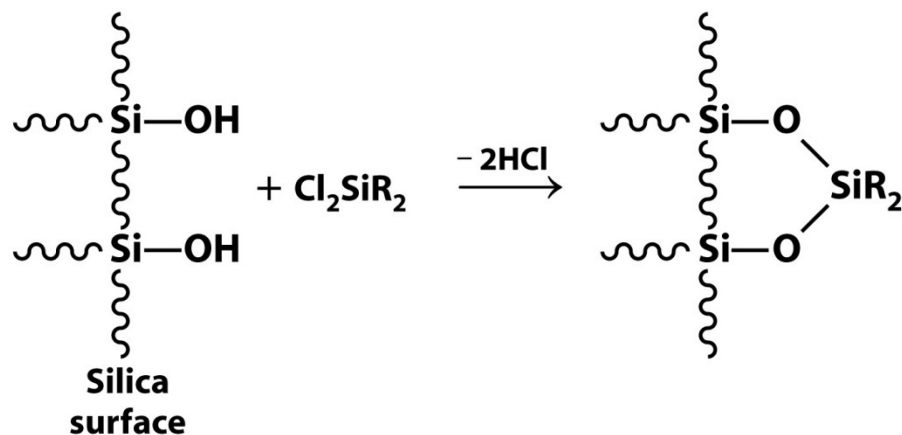
<HPLC system>



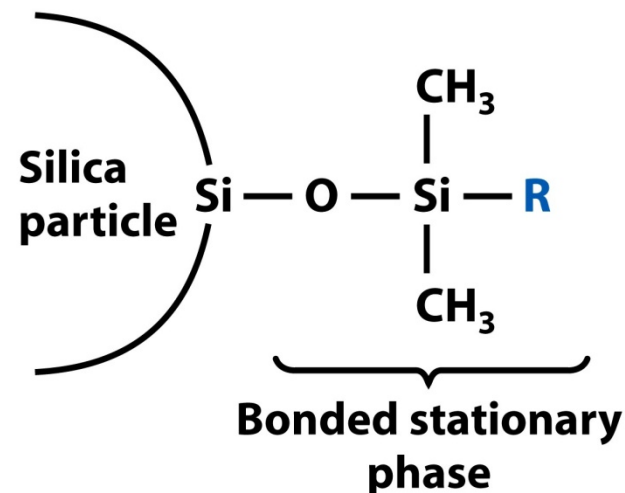
# Bonded Stationary Phase for Partition Chromatography



Unnumbered figure pg 561a  
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Unnumbered figure pg 561b  
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Residual silanol groups on the silica surface are **capped** with trimethylsilyl groups by reacting with  $\text{ClSi}(\text{CH}_3)_3$  to eliminate polar adsorption sites that cause tailing

# Bonded Stationary Phase

**TABLE 24-5** Selection of bonded stationary phases for HPLC

Bonded group	Polarity	Retention mechanisms	Comments
C <sub>18</sub> , C <sub>8</sub> , C <sub>4</sub>	Nonpolar	van der Waals	C <sub>8</sub> does not retain hydrophobic compounds as strongly as C <sub>18</sub>
Phenyl	Nonpolar	Hydrophobic and pi-pi	
Cyano	Intermediate	Hydrophobic, dipole-dipole, and pi-pi	Resolves polar organic compounds by reversed-phase or normal-phase chromatography
Amino	Polar (—NH <sub>2</sub> ) or ionic (—NH <sub>3</sub> <sup>+</sup> )	Dipole-dipole and H-bonding	Normal-phase or ion-exchange separations; separates carbohydrates, polar organic compounds, and inorganic ions; reacts with aldehydes and ketones
Bare silica	Very polar	H-bonding	Normal-phase separations

For a free column selection tool, see <http://www.usp.org/USPNF/columnsDB.html>.

SOURCE: C. S. Young and R. J. Weigand, "An Efficient Approach to Column Selection in HPLC Method Development," *LCGC* **2002**, 20, 464.

Harris, *Quantitative Chemical Analysis*, 8e

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# Detectors in HPLC

- **Ultraviolet detector**: most common
- Refractive index (universal)
- Fluorescence
- Electrochemical
- **Conductivity (ion-exchange C)**
- **Mass spectrometry**
- Chemi-(electrochemi-)luminescence

**TABLE 28-1** Performance of HPLC Detectors

HPLC Detector	Commercially Available	Mass LOD* (typical)	Linear Range † (decades)
Absorbance	Yes	10 pg	3–4
Fluorescence	Yes	10 fg	5
Electrochemical	Yes	100 pg	4–5
Refractive index	Yes	1 ng	3
Conductivity	Yes	100 pg–1 ng	5
Mass spectrometry	Yes	<1 pg	5
FTIR	Yes	1 µg	3
Light scattering	Yes	1 µg	5
Optical activity	No	1 ng	4
Element selective	No	1 ng	4–5
Photoionization	No	<1 pg	4

## UV Detector

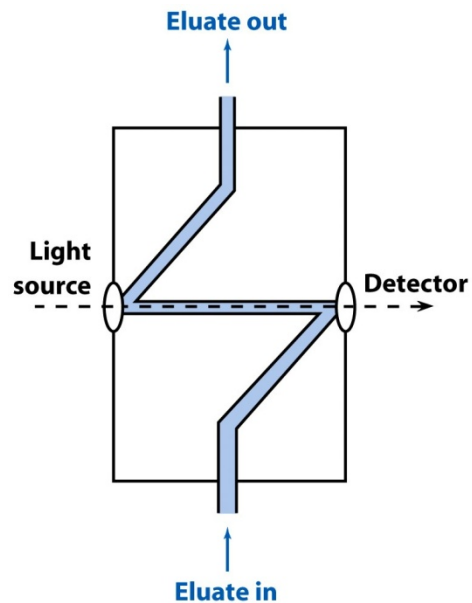


Figure 25-19  
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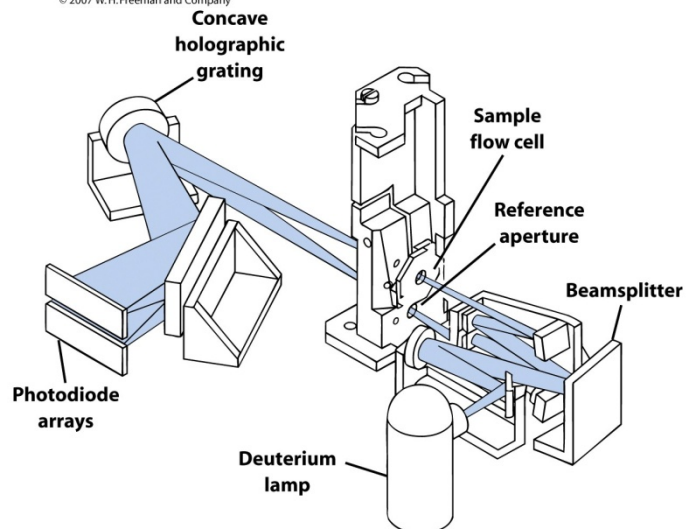


Figure 25-20a  
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## PDA Detector

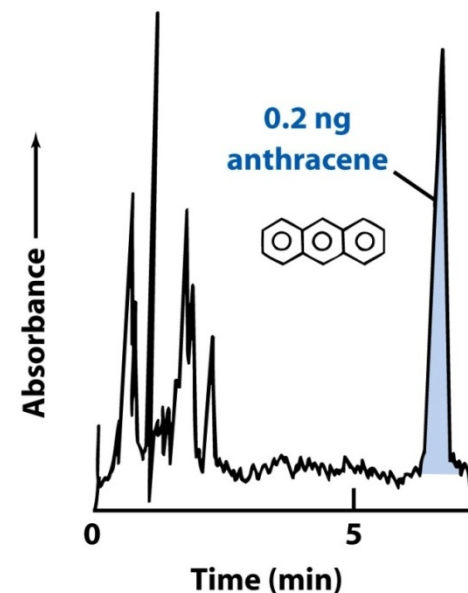


Figure 25-20b  
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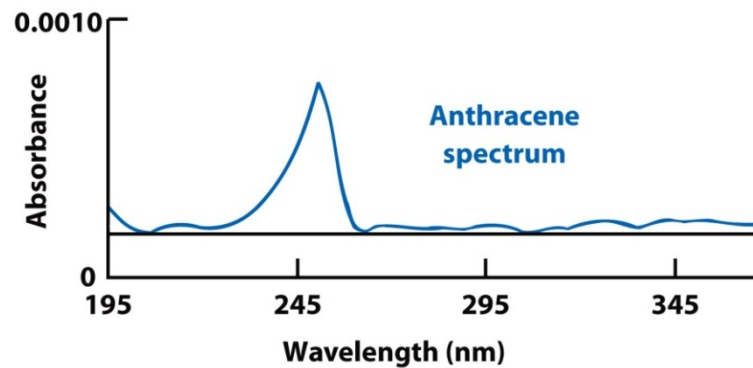
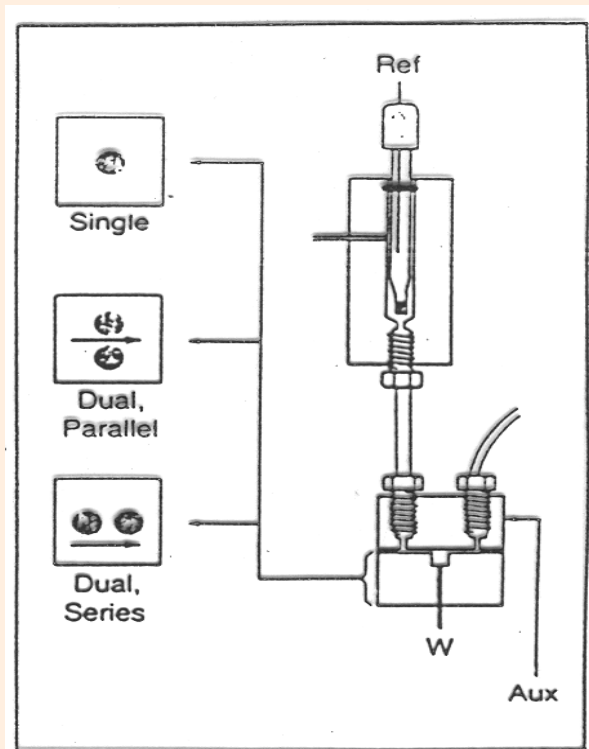


Figure 25-20c  
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## Electrochemical Detector

An electrochemical detector responds to analytes that can be oxidized or reduced such as *phenols, aromatic compounds, peroxides, mercaptans, ketones, aldehydes, conjugated nitriles, aromatic halogen compounds, and aromatic nitro compounds*



## Pulsed electrochemical detection for carbohydrate

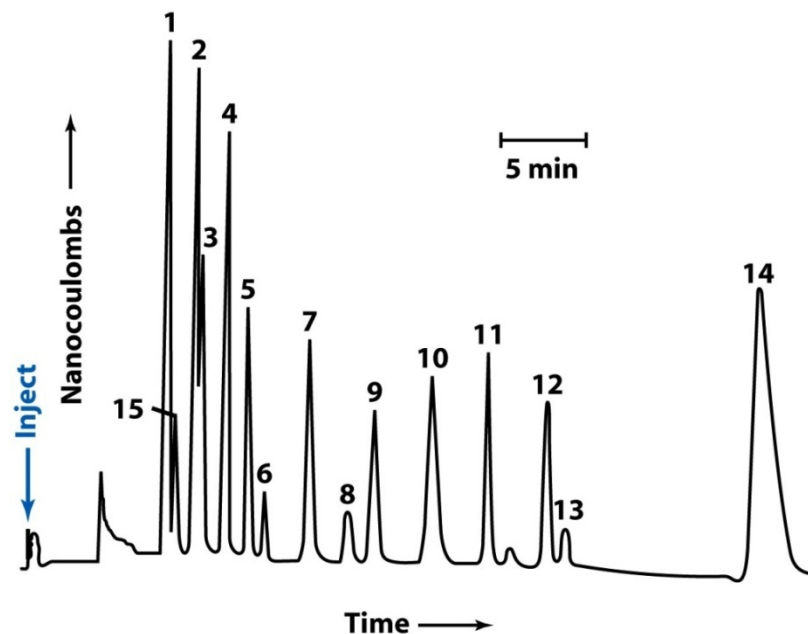


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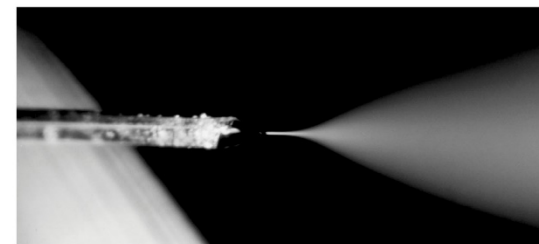
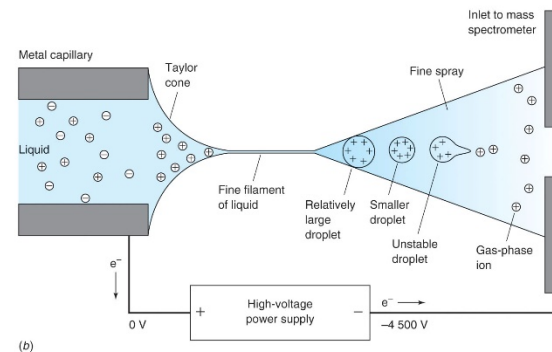
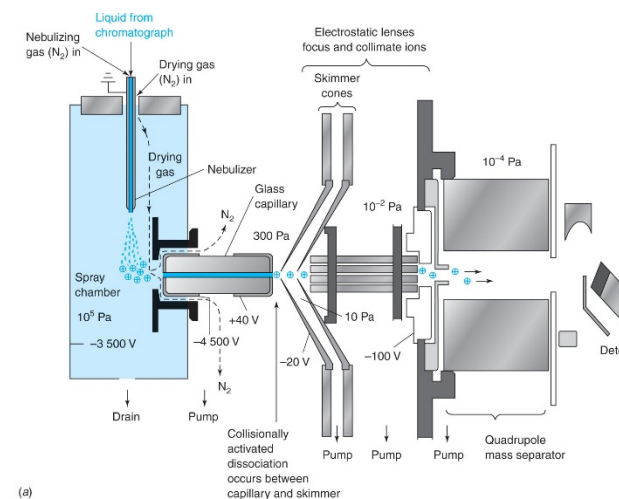


# LC-Mass

Mass spectrometry requires high vacuum to prevent molecular collisions during ion separation

LC creates a huge volume of gas when solvent evaporizes at the interface between column and mass spectrometer

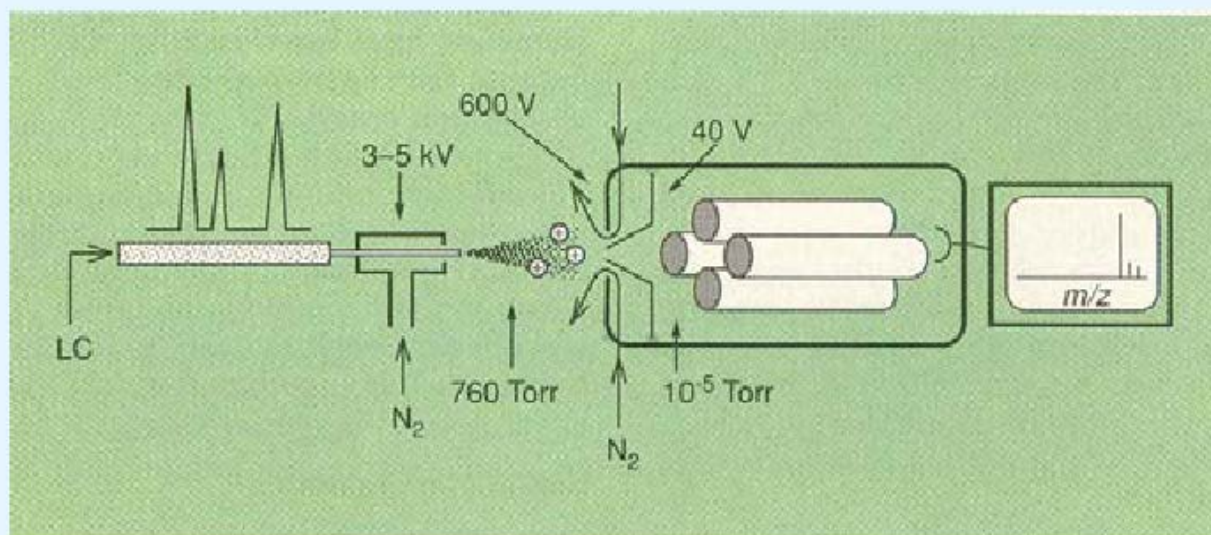
→ Most of this gas must be removed prior to ion separation



**FIGURE 21-21** (a) Pneumatically assisted electrospray interface for mass spectrometry. (b) Gas-phase ion formation. [Adapted from E. C. Huang, I. Wachs, J. I. Conboy, and J. D. Henion, "Atmospheric Pressure Ionization Mass Spectrometry," *Anal. Chem.* 1990, 62, 713A and P. Kebarle and L. Tang, "From Ions in Solution to Ions in the Gas Phase," *Anal. Chem.* 1993, 65, 972A.] (c) Electrospray from a silica capillary. [Courtesy R. D. Smith, Pacific Northwest Laboratory, Richland, WA.]

# Incompatibilities between HPLC and MS

- MS cannot accept HPLC solvent volume (HPLC 500-4,000 mL/min of gas MS pumps about 10-50 mL/min)
- Conventional gas phase ionization in MS not suitable for compounds separated by HPLC which are thermally labile, polar, or high molecular weight

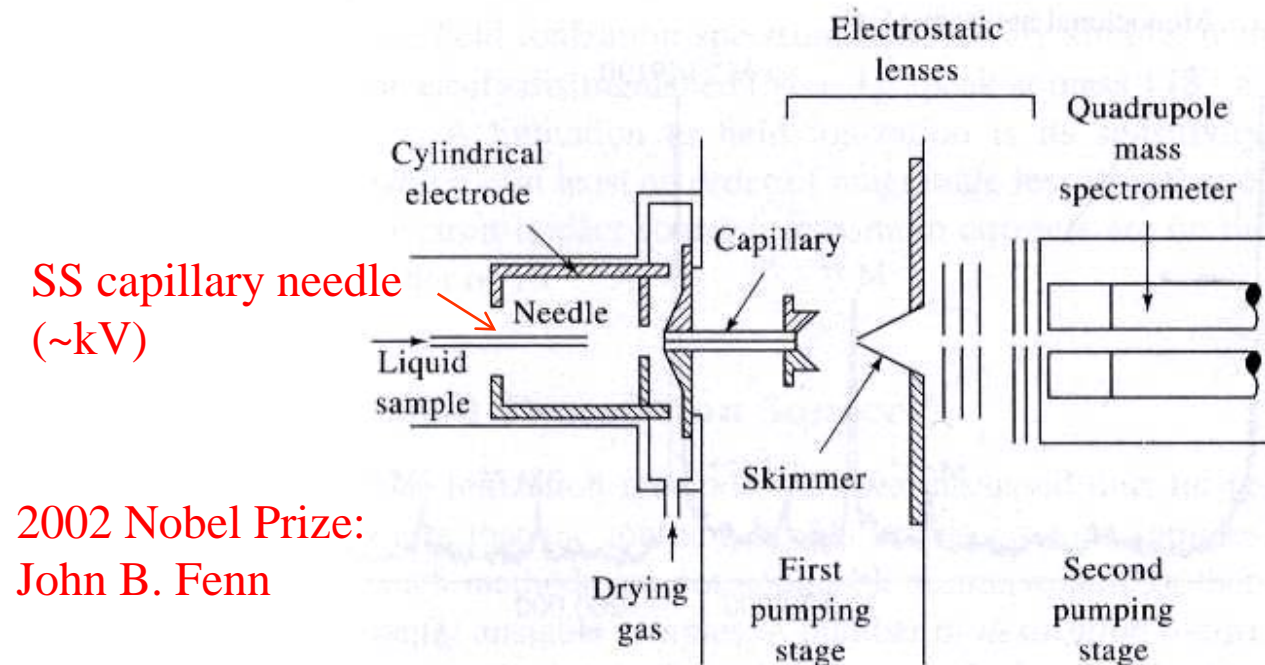


Two most dominant ionization methods in LC-MS

: pneumatically assisted electrospray and atmospheric pressure chemical ionization (APCI)

# Electrospray Ionization

- ESI takes place under atmospheric pressures and temperatures
- Sample is pumped through a SS capillary needle at a rate of few mL/min
- The needle is maintained at  $\sim$  kV against a cylindrical electrode that surround the needle
- Charged spray of fine droplet pass through a desolvating capillary  
→ evaporation of solvent and formation of charged analyte



**Figure 20-8** Apparatus for electrospray ionization. (From J. B. Fenn et al., Science, 1989, 246, 65.)

# **Chapter 30:**

# **Capillary Electrophoresis**

# Electrophoresis

**Electrophoresis:** separation method based on differential rate of migration of charged species in a buffer solution under the influence of an electric field

First developed by the Swedish chemist **Arne Tiselius** in the 1930s: **1948 Nobel Prize**

Analytes: - inorganic anions, cations

- amino acids
- carbohydrates
- peptides, proteins
- nucleic acids, polynucleotides

Special strength of electrophoresis: **separation of charged macromolecules**

- proteins (enzymes, hormones, antibodies)
- nucleic acids (DNA, RNA)

# Basis for Electrophoresis

$$\mathbf{v} = \mu_e \mathbf{E}$$

$v$ : migration velocity of an ion ( $\text{cm s}^{-1}$ )

$\mu_e$ : electrophoretic mobility ( $\text{cm}^2 \text{V}^{-1} \text{s}^{-1}$ )

$E$ : electric field ( $\text{V cm}^{-1}$ )

## Electrophoretic mobility( $\mu_e$ )

- Proportional to the ionic charge on the analytes
- Inversely proportional to frictional retarding force determined by
  - (1) size and shape of the ion
  - (2) the viscosity of the medium in which analyte migrates

For the same size:

The greater the charge  $\rightarrow$  the greater the driving force  $\rightarrow$  faster migration

For the ions of the same charge:

Smaller ion  $\rightarrow$  smaller frictional force  $\rightarrow$  faster migration

Therefore, the ion's charge-to-size ratio determines the electrophoretic mobility

# Capillary Electrophoresis (CE)

## Capillary Electrophoresis:

- Use of fused silica ( $\text{SiO}_2$ ) capillary tube (50 cm long, inner diameter: 25-75  $\mu\text{m}$ )
- Electric field: 30 kV
- High speed, high-resolution separations
- Exceptionally small sample volumes (0.1 – 10 nL)

Migration rate:

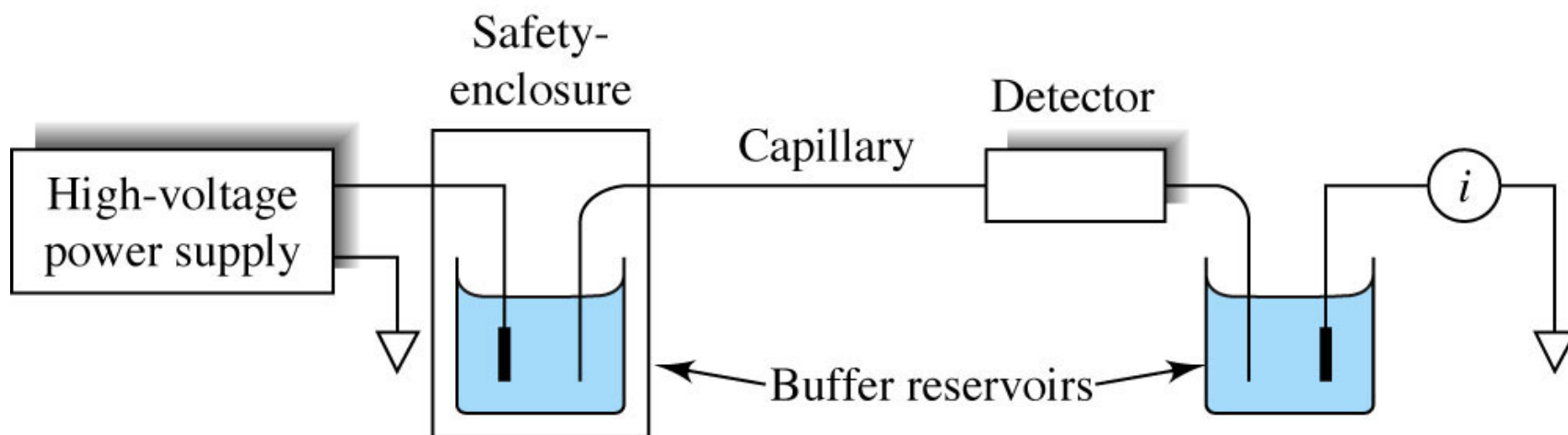
$$v = \mu_e \cdot (V/L)$$

v: migration rate

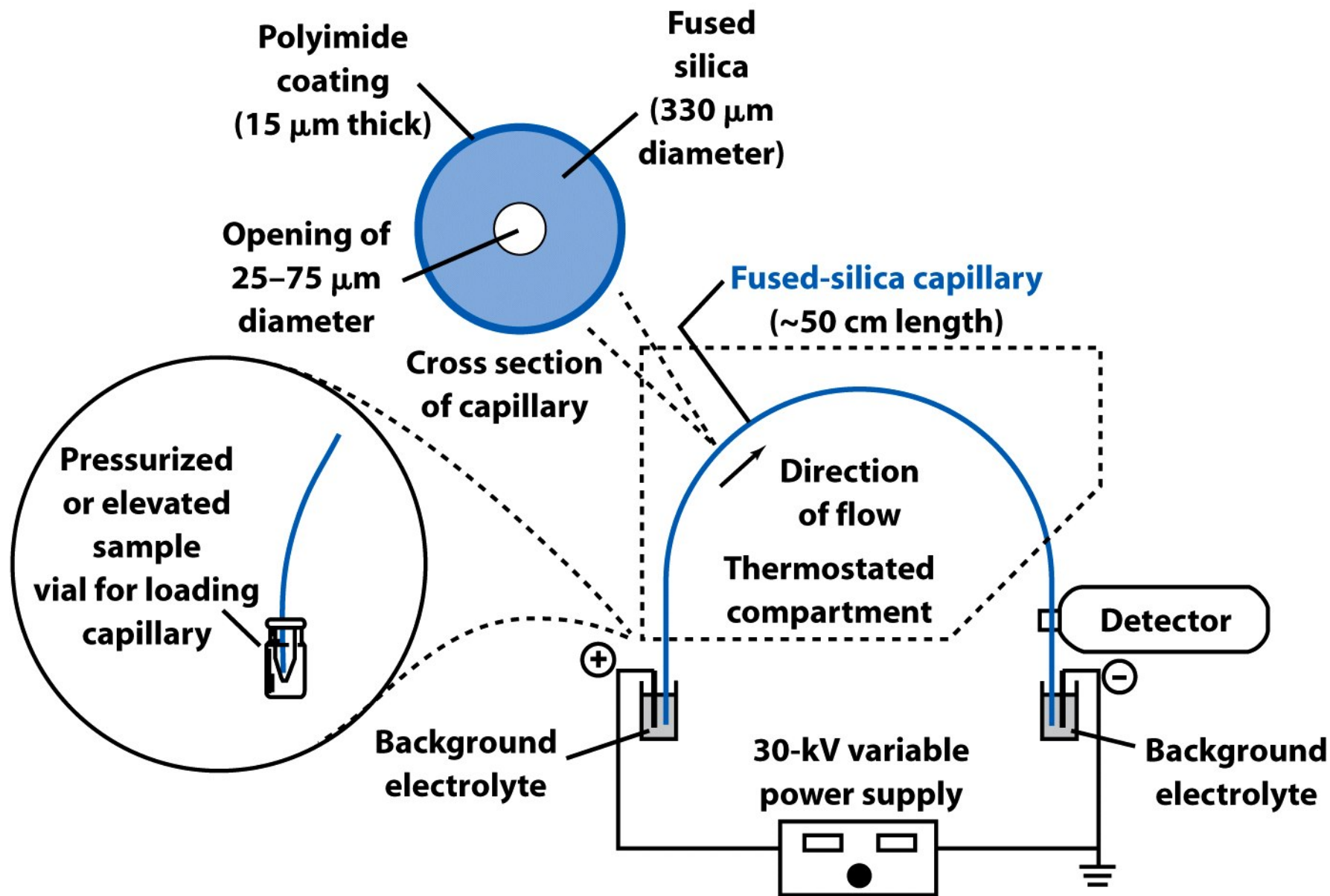
V: applied voltage in volts

L: length over which the voltage  
is applied

Higher voltage → higher separation speed







**Figure 26-17**  
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# Electroosmosis

The inside of a fused silica wall is covered with silanol ( $\text{Si-OH}$ ) groups with a negative charge ( $\text{Si-O}^-$  above  $\text{pH}=2$ )

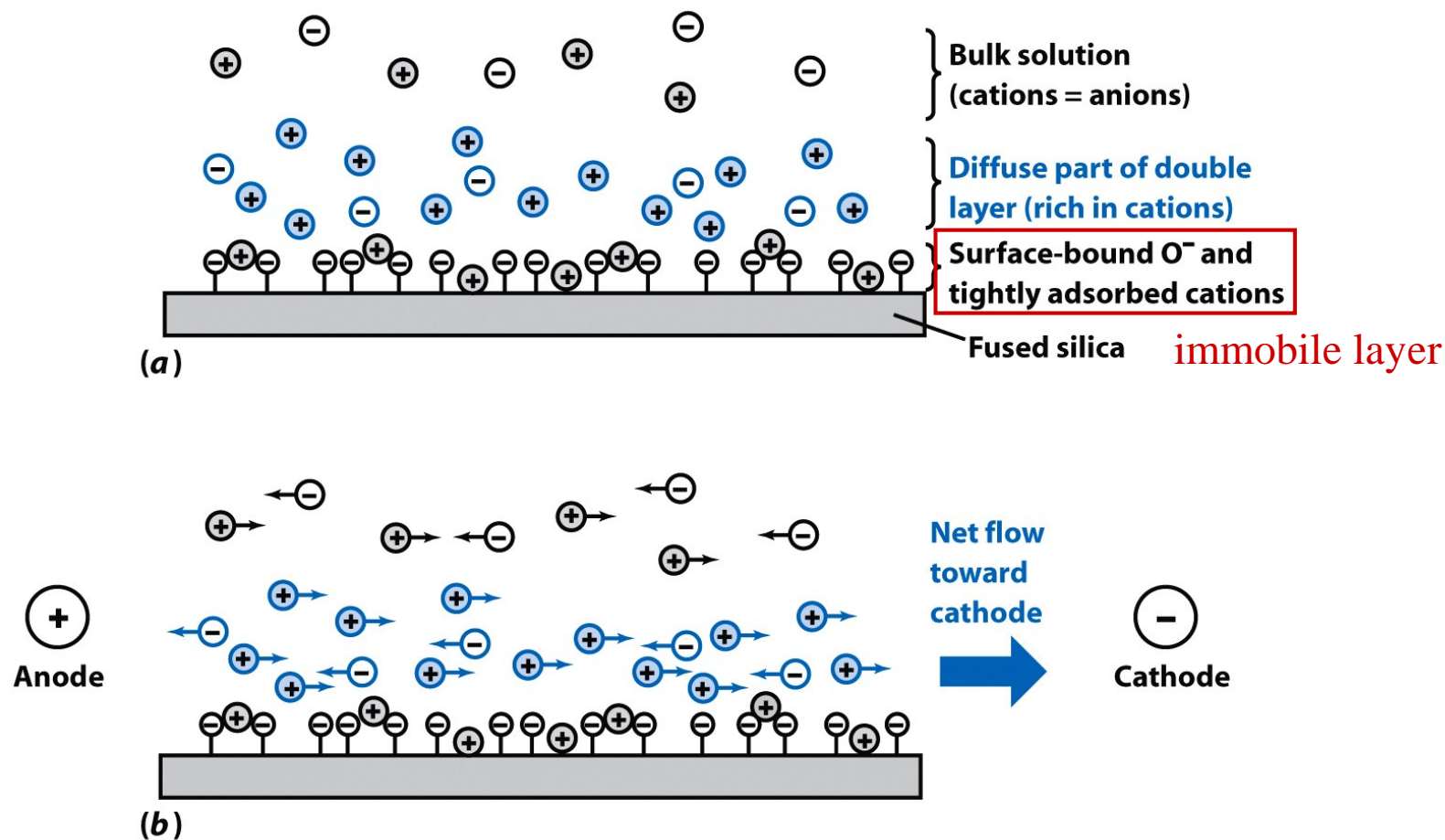


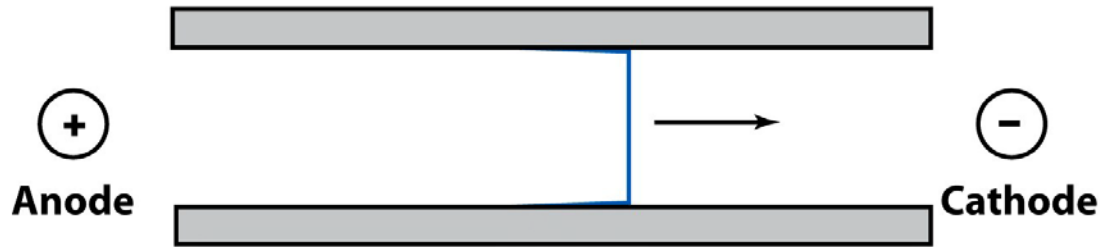
Figure 26-20  
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# Electroosmosis

Electroosmotic velocity:  $v_{eo} = \mu_{eo} \cdot E$

electroosmotic mobility: proportional to surface charge density (higher pH  $\rightarrow$  faster)  
inversely proportional to the square root of ionic strength

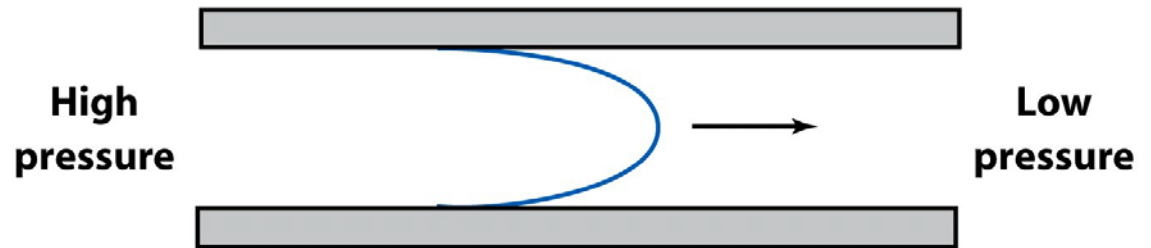
Electroosmotic flow



(a)

Electroosmotic velocity profile: flat flow

Hydrodynamic flow:  
Parabolic velocity profile



(b)

Hydrodynamic velocity profile  
(laminar flow)

# Variations of Capillary Electrophoresis

Capillary Zone Electrophoresis (CZE)

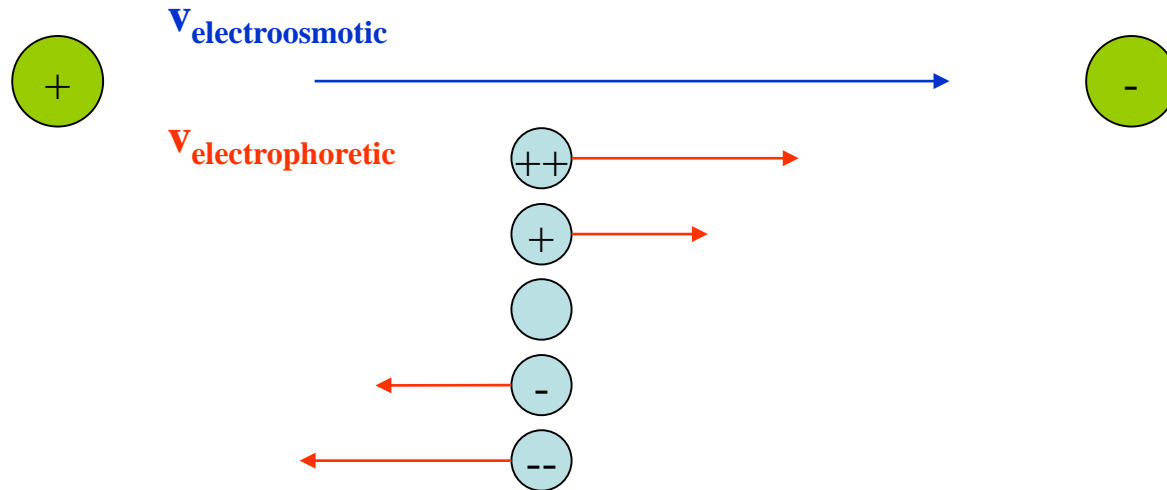
Capillary Gel Electrophoresis (CGE)

Capillary Electrochromatography

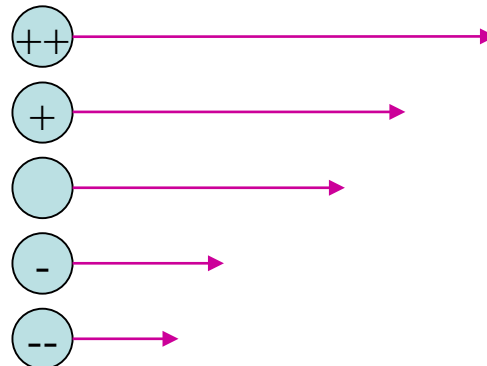
Micellar Electrokinetic Capillary Chromatography (MECC)

Capillary Isoelectric Focusing

# Capillary Zone Electrophoresis



$$V_{\text{total}} = V_{\text{electroosmotic}} + V_{\text{electrophoretic}}$$



# Capillary Gel Electrophoresis

Macromolecules are separated in a gel by **sieving**:

Smaller molecules → migrate faster than larger molecules through gel

Analytes: proteins, DNA fragments, oligonucleotides)

Qualitative analysis can be conducted by comparing the patterns produced to standards.

This example is a molecular weight determination of proteins but other materials can be evaluated.

This approach is used in genetic 'fingerprinting.'

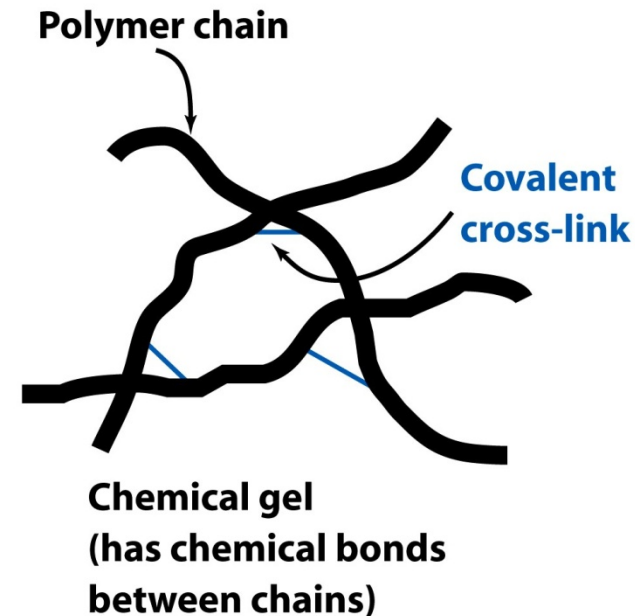
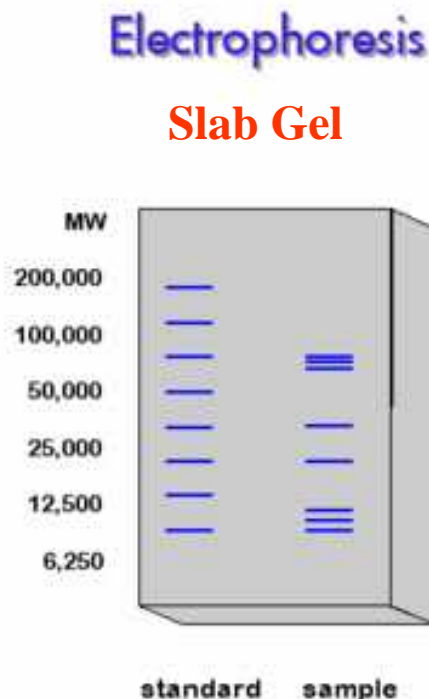


Figure 26-36a  
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Polyacrylamide gel

# Capillary Gel Electrophoresis

- Proteins are first denatured by reducing their disulfide (-S-S-) bonds with excess 2-mercaptoethanol and adding SDS( $\text{C}_{12}\text{H}_{25}\text{OSO}_3^-\text{Na}^+$ )
- Dodecyl sulfate anion coats hydrophobic regions and gives the protein a large negative charge that is approximately proportional to the length of protein

**Sodium dodecyl sulfate  
(SDS)- Capillary gel  
electrophoresis**

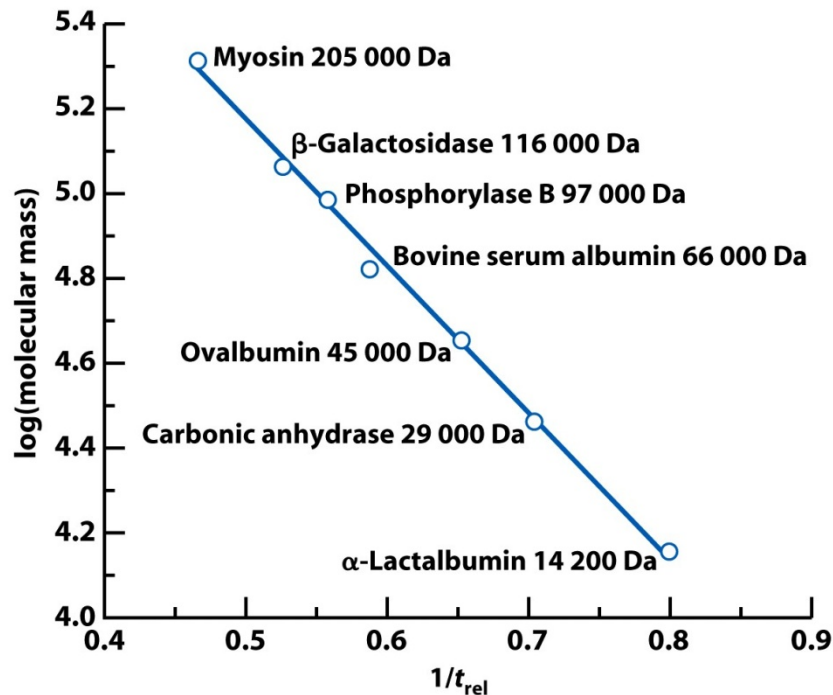


Figure 26-38  
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# Detectors in Capillary Electrophoresis

Highly sensitive detector is required in CE (extremely small sample amount)

- UV detector : most general
- Fluorescence detector (laser-induced fluorescence): good sensitivity
- Amperometric detector
- Conductivity detector
- Chemiluminescence detector
- Mass spectrometer



# Detectors in Capillary Electrophoresis

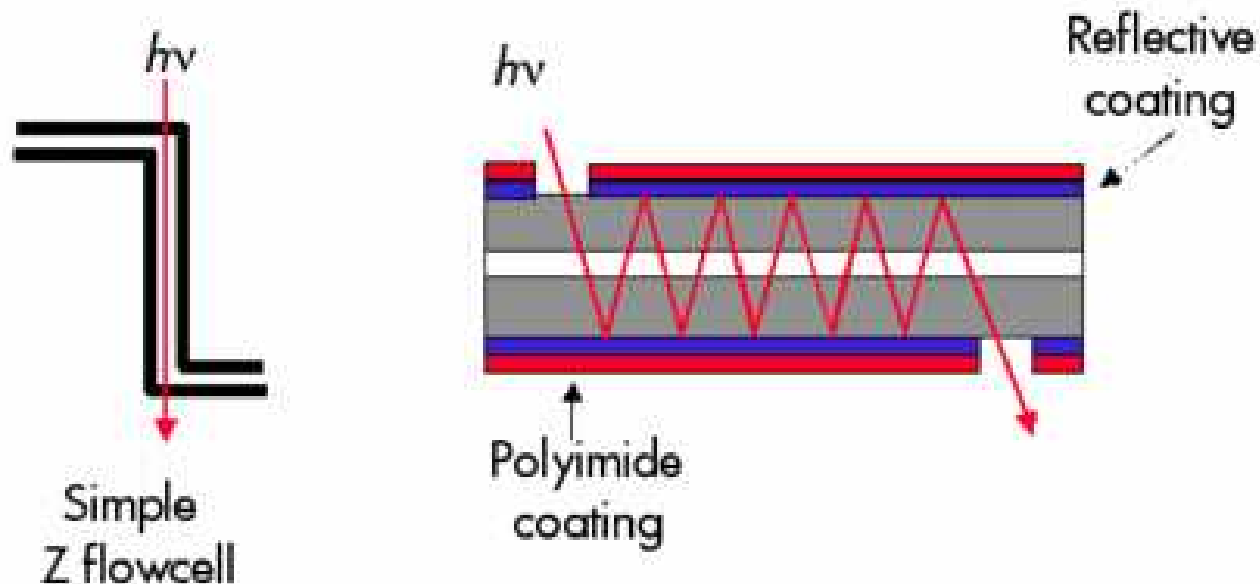
**TABLE 30-1** Detectors for CE

Type of Detector	Representative Detection Limit* (attomoles detected)
Spectrometry	
Absorption <sup>†</sup>	1–1000
Fluorescence	1–0.01
Thermal lens <sup>†</sup>	10
Raman <sup>†</sup>	1000
Chemiluminescence <sup>†</sup>	1–0.0001
Mass spectrometry	1–0.01
Electrochemical	
Conductivity <sup>†</sup>	100
Potentiometry <sup>†</sup>	1
Amperometry	0.1

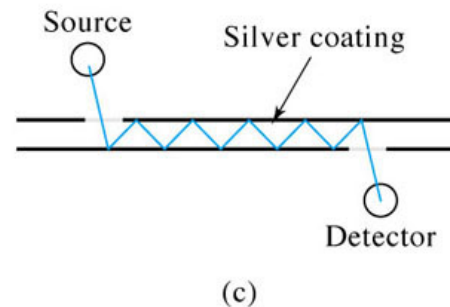
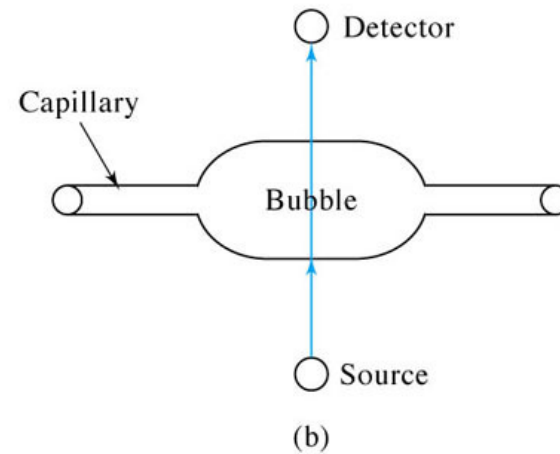
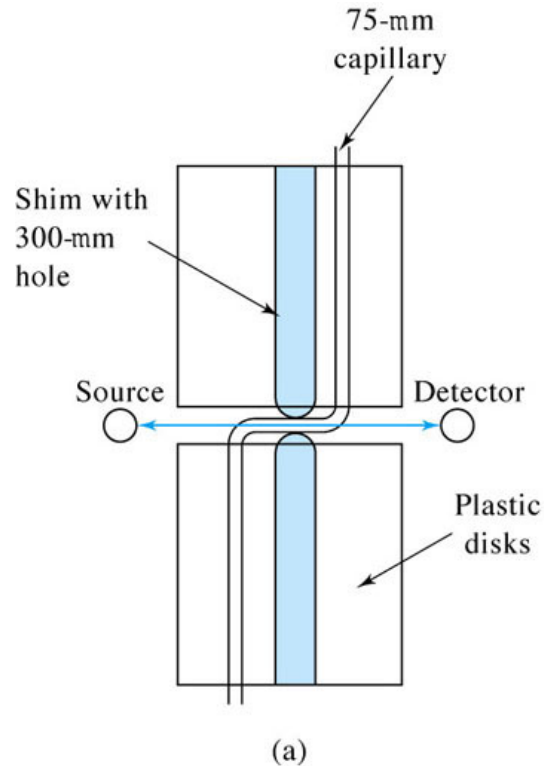
# Detectors in Capillary Electrophoresis

UV/Vis

Because of the small volumes, one must get 'creative' to obtain a measurable response -- while still in the capillary tube.



# Detectors in Capillary Electrophoresis



# **Detection of Attomolar Concentrations of Alkaline Phosphatase by Capillary Electrophoresis Using Laser-Induced Fluorescence Detection**

**Douglas B. Craig, Jerome C. Y. Wong, and Norman J. Dovichi\***

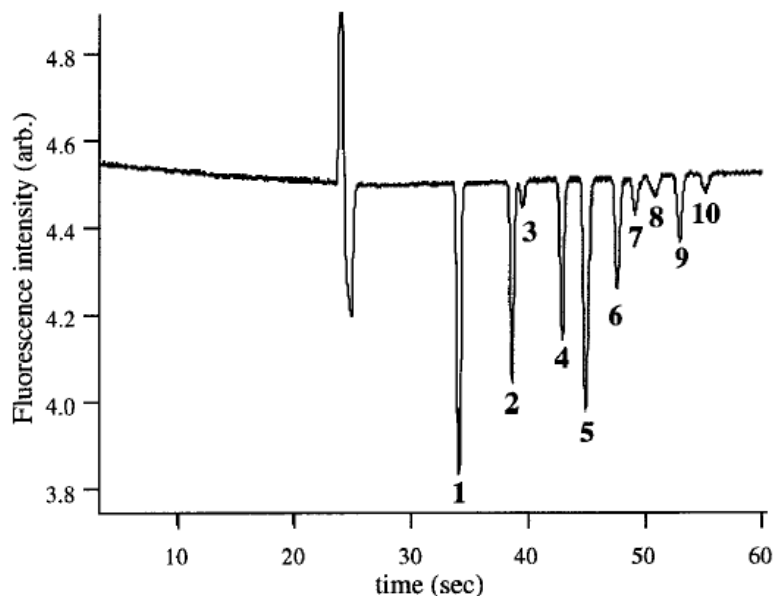
*Department of Chemistry, University of Alberta, Edmonton, Alberta T6G 2G2, Canada*

Alkaline phosphatase can be assayed by monitoring the conversion of the fluorogenic substrate AttoPhos into the highly fluorescent product AttoFluor. We have used capillary electrophoresis with laser-induced fluorescence detection to monitor this reaction. The concentration limit of detection ( $3\sigma$ ) of alkaline phosphatase is  $1.5 \times 10^{-17}$  M (2.1 fg/mL), which corresponds to a mass limit of detection of nine molecules ( $1.5 \times 10^{-23}$  mol) contained within a 1- $\mu$ L sample volume.

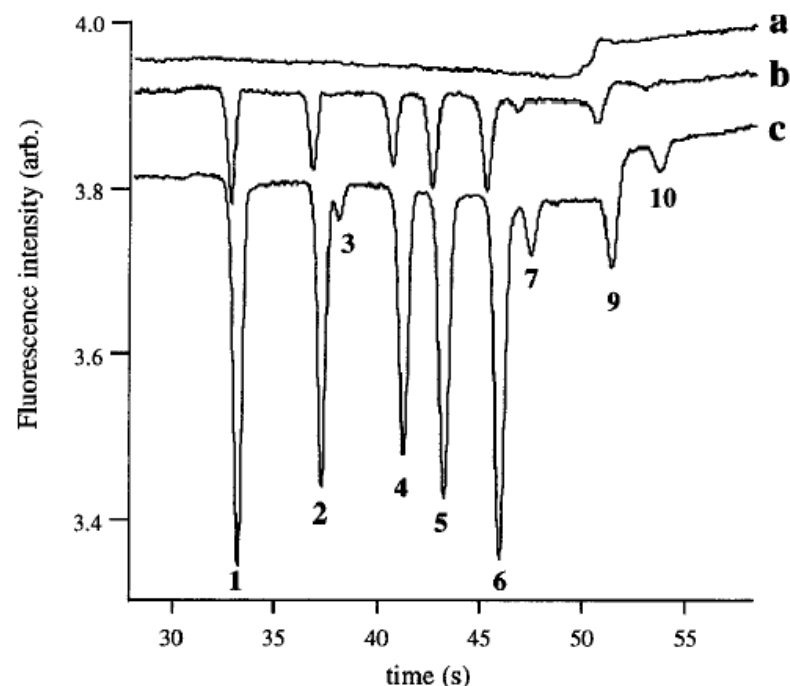
# Separation and Detection of Explosives on a Microchip Using Micellar Electrokinetic Chromatography and Indirect Laser-Induced Fluorescence

Susanne R. Wallenborg and Christopher G. Bailey\*

Sandia National Laboratories, P.O. Box 969/ms9671, Livermore, California 94551



**Figure 1.** Chip-based MEKC-IDLIF electropherogram of the EPA 8330 mixture of nitroaromatics and nitramines. Analytes: 20 ppm of each TNB (1), DNB (2), NB (3), TNT (4), tetryl (5), 2,4-DNT (6), 2,6-DNT (7), 2-, 3-, and 4-NT (8), 2-Am-4,6-DNT (9), and 4-Am-2,6-DNT (10). Conditions: MEKC buffer, 50 mM borate, pH 8.5, 50 mM SDS, 5  $\mu$ M Cy7, separation voltage 4 kV, separation distance 65 mm.



**Figure 7.** MEKC-IDLIF analysis of extracts from spiked soil samples: soil blank (a), soil containing 1 ppm of each analyte (b), and soil containing 5 ppm of each analyte (c). Peak identifications and conditions were as for Figure 1.

# Detectors in Capillary Electrophoresis

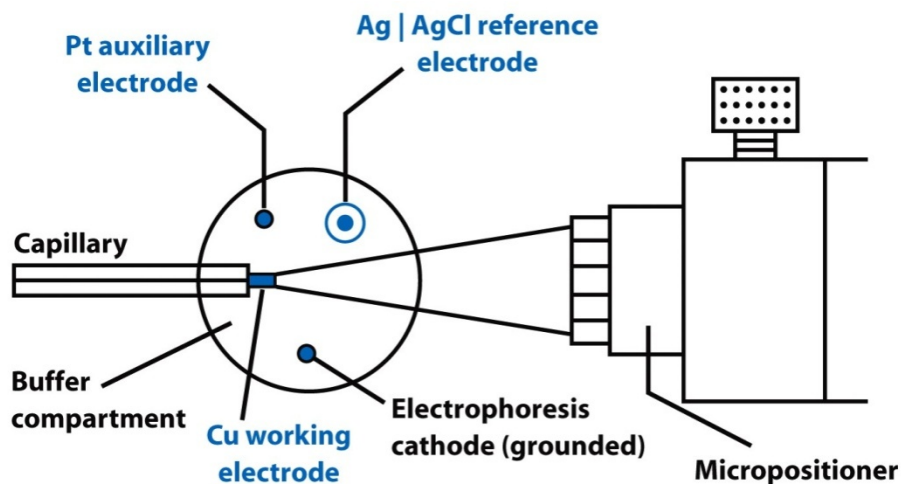


Figure 26-29a  
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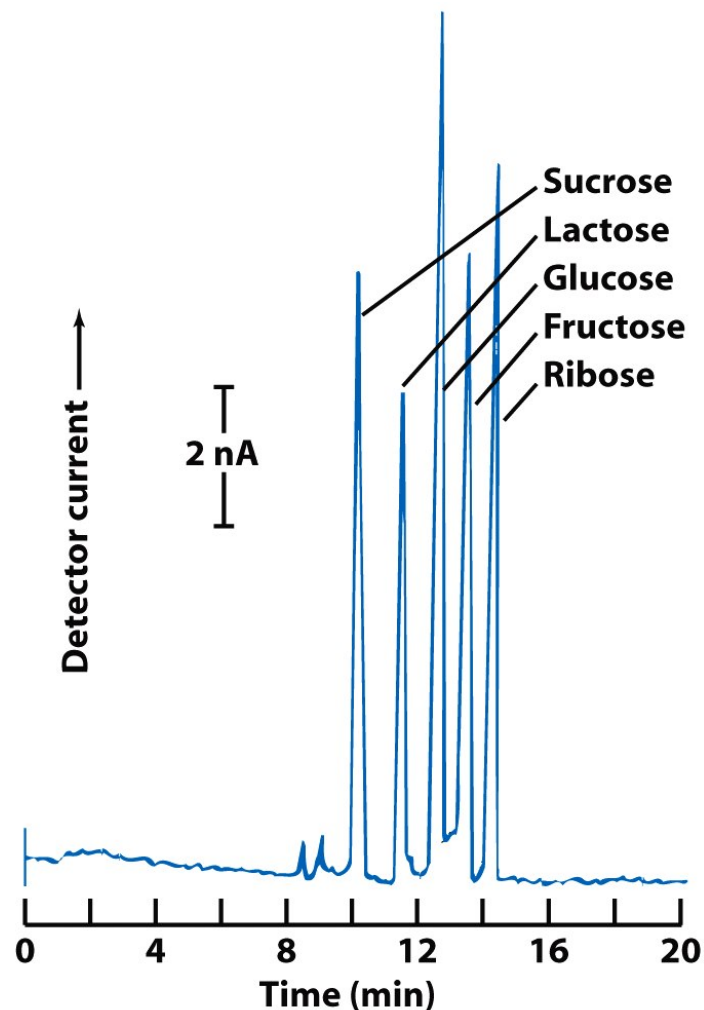
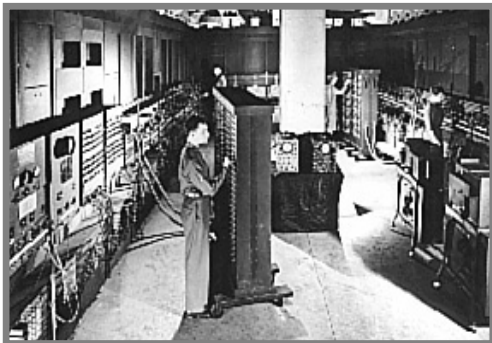
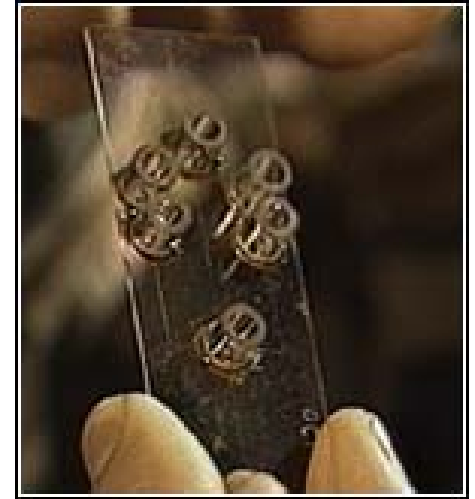
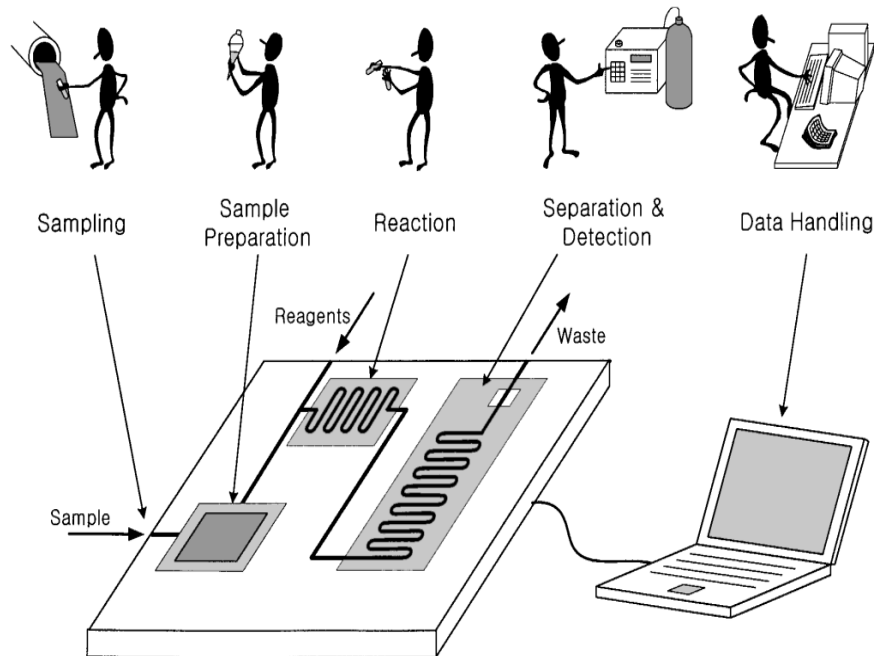


Figure 26-29b  
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# Lab-on-a-Chip



# Lab-on-a-Chip



(CNN)

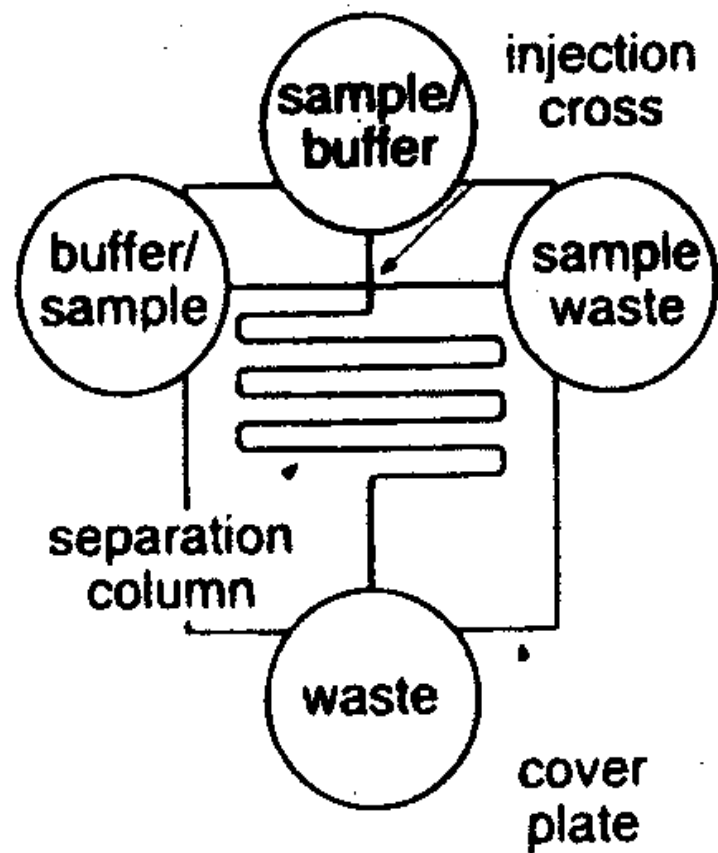
D. J. Harrison, et. al., *Science*, **1993**, 261, 895-897

- Highly integrated system (sample pretreatment, reaction, separation, and detection all on-a-chip)
- High speed analysis (few second or few minutes)
- Ultra-low volume - minimal reagent or sample consumption (1-10 pL)
- Highly parallel - many samples at once: high-throughput
- Small and possibly portable system
- Potentially disposable assay systems



# Lab-on-a-Chip

Agilent 2100



DNA  
RNA  
Protein